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(54) Title: HUMAN LACTOFERRIN (57) Abstract The present invention relates to a human lactoferrin cDNA gene obtained from human breast tissue and the protein encoded therefrom. The present invention further relates to methods for detecting malignancy arising from tissues that normally secrete lactoferrin using the cDNA gene probe of the present invention. Another aspect of the present invention relates to the promotor region that regulates the human lactoferrin gene.		

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HUMAN LACTOFERRIN

BACKGROUND OF THE INVENTIONFIELD OF THE INVENTION

The present invention relates to a human lactoferrin gene isolated from breast tissue and to the protein product encoded therein. The present invention further relates to the promotor region of human lactoferrin gene and to methods for detecting and analyzing malignancies arising from tissues that normally secrete lactoferrin using a novel human lactoferrin cDNA gene sequence.

BACKGROUND INFORMATION

Lactoferrin is a single polypeptide molecule (M, 76,000) with sites where two oligosaccharide chains can attach (B.F. Anderson et al., *J. Mol. Biol.* 209:711-734 (1989)). This protein shares significant homology with transferrin, however, its role in iron transport is limited since it binds iron 260 times stronger than transferrin (B.F. Anderson et al., (1989)). Two and possibly three isoforms of lactoferrin have been isolated using an affinity chromatography (P. Furnamski et al., *J. Exp. Med.* 170:415-429 (1989); A. Kijlstra et al., *Current Eye Res.*, 8:581-588 (1989)). Lactoferrin has been shown to inhibit bacterial growth by chelating iron and directly attacking the cell wall (R.T. Ellison et al., *Infect Immun.*, 56:2774-2781 (1988)), contribute to the anemia of chronic disease (Birgens. *Scand. J. Haematol.*, 33:225-230 (1984)), improve intestinal absorption of iron in infants (Birgens., (1984)) inhibit myelopoiesis (H.E. Broxmeyer et al., *Blood Cells*

13:31-48 (1987)), and degrade mRNA (P. Furmanski et al., (1989); M.R. Das et al., *Nature* 262:802-805 (1976); P. Furmanski and Z.P. Li, *Exp. Hematol* 18:932-935 (1990). Large quantities of lactoferrin are
5 found in breast milk (B. Lonnerdal et al., *Nutrition Report Int.*, 13:125-134 (1976)), in estrogen-stimulated uterine epithelium (B.T. Pentecost and C.T. Teng, *J. Biol. Chem.* 262:10134-10139 (1987)), and in neutrophilic granulocytes (P.L. Masson et al., *J. Exp. Med.*, 130:643-
10 658 (1969)) with smaller amounts in tears, saliva, serum, and seminal fluid (D.Y. Mason and C.R. Taylor, *J. Clin. Path.*, 31:316-327 (1978)).

While normal breast ductal epithelium and neutrophilic granulocytes contain lactoferrin, their
15 malignant counterparts frequently do not (C. Charpin et al., *Cancer*, 55:2612-2617 (1985); T.A. Rado et al., *Blood*, 70:989-993 (1987)). This has been evaluated at the protein level and in a few samples at the messenger RNA level (T.A. Rado et al., (1987)).
20 Analysis at the genomic level has not been performed. DNA variations, that are detected in the coding regions, may lead to abnormal protein structure and loss of normal function. Variations, such as mutations, deletions, or changes in
25 methylation, at the promoter regions could lead to altered regulation of the gene. Evaluation of the lactoferrin gene may provide interesting insight concerning the production of lactoferrin in malignant cells. Thus, the need exists for the
30 structure of the lactoferrin gene including the cDNA and the promotor region. The present invention provides such a description of the structure of a

human lactoferrin cDNA and promotor region of the gene.

Using a lactoferrin cDNA clone isolated from human breast tissue, the applicants have
5 evaluated restriction fragment length changes in DNA from the white blood cells of 10 normal controls, acute non-lymphocyte leukemia (ANLL) cells from 7 patients, T-cell acute lymphocyte leukemia (ALL) from one patient, 3 leukemia cell lines, and 7
10 breast cancer cell lines. A comparative study of the lactoferrin gene in these different cell types is provided herein.

The present invention further relates, in part, to a human lactoferrin cDNA and the protein
15 product encoded therein. In another aspect, the present invention relates to methods for detecting malignancy in tissues that normally secrete lactoferrin by evaluating restriction patterns in DNA using a lactoferrin gene probe of the present
20 invention.

SUMMARY OF THE INVENTION

It is an object of the present invention to provide a DNA sequence of the human lactoferrin gene including the cDNA and the promotor region and
25 to the protein product encoded therein.

In one embodiment, the present invention relates to a DNA segment encoding human lactoferrin according to the sequence identification number In another embodiment, the present invention relates to
30 the human lactoferrin protein encoded by the sequences given in identification number 2.

In yet another embodiment, the present invention relates to a DNA segment of the promotor region for human lactoferrin according to the sequence identification number 5 and allelic variations thereof.

In a further embodiment, the present invention relates to a recombinant DNA construct comprising the DNA segments encoding the human lactoferrin gene sequences described above and a vector.

In another embodiment, the present invention relates to a recombinant DNA construct comprising the DNA segment encoding the human lactoferrin gene described above and a DNA promotor regulatory region for human lactoferrin according to sequence identification number 5 or portion thereof operatively linked to the DNA fragment.

In a further embodiment, the present invention relates to a host cell comprising the above described constructs.

Another embodiment of the present invention relates to a method of treating a condition in a patient characterized by a deficiency in lactoferrin by administering to the patient an amount of human lactoferrin according to the present invention in sufficient quantities to eliminate the deficiency. The conditions include neutropenia, AIDS, skin infection, gastrointestinal bacterial overgrowth syndrome, vaginal infection and septic shock.

In yet another embodiment, the present invention relates to methods of diagnosing malignancy or detecting the recovery of a malignancy

from a biological sample comprising the steps of isolating DNA from the biological sample and from normal control samples, cutting the DNA with a restriction enzyme called Xba I, hybridizing the cut
5 DNA with a DNA segment of the human lactoferrin gene of the present invention described above or portion thereof under conditions such that hybridization is effected and comparing the hybridization product patterns of the biological sample and the normal
10 control sample with each other.

In a further embodiment, the present invention relates to a method for detecting small insertions, deletions or mutations surrounding the human lactoferrin gene comprising the steps of
15 isolating the DNA from a biological sample suspected of having such an insertion, deletion or mutation, amplifying the DNA using the human lactoferrin gene segment of the present invention described above or portion thereof in a polymerase chain reaction
20 followed by enzymatically cutting the amplified DNA with Xba I, and hybridizing this DNA with the human lactoferrin gene segment described above under conditions such that hybridization is effected and sequencing the hybridized DNA.

25 Various other objects and advantages of the present invention will become obvious from the drawings and detailed description of the invention.

The entire contents of all publications mentioned herein are hereby incorporated by
30 reference.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the immunocytochemical staining of normal bone marrow (A) x 400, and breast cancer cell line SKB R3 (B) x 680 using anti-lactoferrin antibody at 1:1500.

5 Figure 2 depicts the restriction fragments produced with DNA from normal cells (A) or from leukemia cells (B) using lactoferrin cDNA (HLF 1212) as the probe. Normal samples (n=9) and DNA from 10 different leukemia cells types were digested with indicated enzyme, run in one gel and representative lanes cut out for comparison.

15 Figure 3 depicts the restriction fragments produced using DNA from normal samples (A) and from breast cancer cell lines (B), using lactoferrin cDNA (HLF 1212) as a probe. Normal samples (n=2) and DNA from eight cancer lines were digested with indicated enzyme, run in the same gel, and representative lanes cut out for comparison.

20 Figure 4 shows the restriction fragments produced using Msp I and lactoferrin cDNA (HLF 1212) as the probe. Lanes 1 - 9 are DNA from normal donors. Lanes 10 - 16 represent DNA from leukemia cells from patients. Lane 17 is cell line K562, lane 18 is KG 1, and lane 19 is U937.

25 Figure 5 represents the restriction fragments produced using Msp I and lactoferrin cDNA (HLF 1212) as the probe. Lanes 1 and 2 are DNA from normal donors. Lanes 3 - 9 represent DNA from breast

cancer cell lines. The cell lines are in the following order: Lane 3 - MDAMB 468, lane 4 - MCF 7, lane 5 - BT 474, lane 6 - HBL 100, lane 7 - MDA 175, lane 8 - SKB R3, lane 9 - ZR 75-1.

5 Figure 6 shows the restriction fragments produced using Xba I and lactoferrin cDNA (HLF 1212) as the probe. Lanes 1 - 9 are DNA from normal donors. Lanes 10 - 16 are DNA from leukemia cells from patients and lanes 17 - 19 DNA from leukemia cell
10 lines (lane 17 - K562, lane 18 - KG1, lane 19 - U937). Arrow A is the band found in patterns A (lanes 1, 2, and 7), B, and C. Arrow B is the band found in patterns B (lanes 3 - 6, 8 - 10, 13, 14) and C. Arrow C is only found in pattern C (lanes 11, 12, 16). Insert is the same specimens run on a 0.7%
15 agarose gel.

Figure 7 depicts the restriction fragments produced using Xba I and lactoferrin cDNA (HLF 1212) as the probe. Lanes 1 and 2 are DNA from normal donors.
20 Lanes 3 - 9 are DNA from breast cancer cell lines. The order is: Lane 3 - MDAMB 468, lane 4 - BT 474, lane 5 - HBL 100, lane 6 - MDA 175, lane 7 - SKB R3, lane 8 - ZR 75-1, lane 9 - ZR 75-30. Restriction fragment patterns as discussed in the text are in
25 the following lanes: pattern A is seen in lane 1, pattern B in lane 2, and pattern D in lanes 3 - 9.

Figure 8 shows the restriction fragments produced using Hpa II and lactoferrin cDNA (HLF 1212) as the probe. Lanes 1 - 9 are DNA from normal donors.
30 Lanes 10 - 16 are DNA from leukemia cells from

patients. Lane 17 is cell line KG1, lane 18 is U937, and lane 19 is HL 60.

Figure 9 shows the restriction fragments produced using Hpa II and lactoferrin cDNA (HLF 1212) as the probe. Lanes 1 and 2 are DNA from normal donors. Lanes 3 - 10 are breast cancer cell lines in the following order: lane 3 - MDAMB 468, lane 4 - MCF 7, lane 5 - BT 474, lane 6 - HBL 100, lane 7 - MDA 175, lane 8 - SKB R3, lane 9 - ZR 75-1, lane 10 - ZR 75-30.

Figure 10 depicts a sequence data of HLF 1212. Differences between the published protein derived AA sequence and our cDNA derived sequence are indicated by underlining the extra AA in our sequence or indicating substitutions beneath our sequence. Nucleotide differences based on published sequence data are indicated above our sequence. Nucleotide changes resulting in a different AA are typed below the area of substitution.

20 DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to a cDNA sequence for human lactoferrin and the protein encoded therein. The cDNA called HLF1212 was isolated from human breast tissue and is 2117 kb in length. The sequence agrees with the modified amino acid sequence of iron-binding lactoferrin in all areas except the 3 sites in the N-terminal region. One further change is in arginine in place of a lysine at amino acid 200.

30 Another aspect of the present invention relates to methods for diagnosing malignancy by

restriction fragment length polymorphisim (RFLP) analysis of DNA extracted from normal peripheral blood and leukemia cells from patients using the cDNA of the present invention as the probe.

5 Southern analysis indicates that the human lactoferrin gene is polymorphic when tested using Msp I and Xba I restriction enzymes. Further analysis indicates that the changes in the XbaI recognition site could be explained by alterations
10 in DNA caused by or resulting in malignancy. In the present invention, the DNA from normal and malignant cells are digested with XbaI and the fragment pattern compared using methods well known in the art. The Xba I restriction is associated with 4
15 patterns in normal and malignant cells (Example 3 and Figures 6 and 7). The most striking change is the deletion of many bands found only in DNA obtained from malignant cells or cell lines derived from either leukemia or breast cancer.

20 If the patterns found in Example 3 (Xba I RFLP pattern C + D) are found in women before breast cancer occurs, it may be easy to screen women at high risk of breast cancer for these changes using cDNA probe of the present invention and RFLP
25 methodologies well known in the art. For example, lymphocytes may be separated from peripheral blood, DNA extracted, and cut with XbaI. This DNA can then be probed with HLF 1212 or a small piece of HLF 1212 and patterns determined. High risk patients may be
30 placed on preventive medicines such as Tamoxifen retinoids or have surgery. The same may hold for other hormonally responsive tumors such as prostate, uterus, or tumors arising from

lactoferrin secreting organs such as leukemia, or salivary gland.

Another aspect of the present invention relates to RFLP methods to measure the prognosis of certain types of cancer patients that are given
5 therapeutics. One may place patients with breast, prostate, uterine, or salivary cancer into risk groups. Those with a specific pattern may be at different risks of disease reoccurrence. Thus, RFLP
10 analysis using the cDNA probe of the present invention may provide prognostic information for patients with cancer.

Another aspect of the present invention relates to methods for detecting small insertions, deletions or mutations surrounding the human
15 lactoferrin gene. Either of the above described RFLP methods could be combine with polymerase chain reaction (PCR) analysis. The abnormal area of the gene may be amplified using methods well known in
20 the art and then mutations detected using restriction analysis (i.e. Xba I) and sequencing.

Yet another aspect of the present invention relates to methods for detecting tumors in pathological specimens that may contain too few
25 malignant cells to be detected by standard methods. This method may involve PCR of DNA extracted from specimens (biopsy of tissue or bone marrow) and subsequent analysis using the RFLP techniques and DNA probes described above and in the Examples.

30 In another embodiment, the present invention relates to the cDNA clone for human lactoferrin called HLF 1213 and the protein encoded therein. The sequence of HLF 1213 (sequence ID

NO:3) is a combination of clones HLF 1212 (sequence ID NO: 1), 031A (sequence ID NO: 5) and other clones isolated in the same method as HLF 1212. (See Example 2). This clone is a composite of the
5 complete human lactoferrin cDNA. This clone may be constructed by splicing 2 clones together with HLF 1212 (031A, and HLF 1212). Both HLF 1212 or this combined fragment called HLF 1213 may be used to make recombinant human lactoferrin.

10 In another embodiment, the present invention relates to the human lactoferrin protein obtained from HLF 1212 and HLF 1213 called sequence ID Numbers 2 and 4 respectively.

15 In yet another embodiment, the present invention relates to recombinant human lactoferrin expressed in vitro through molecular genetic engineering technology.

The present invention also relates to the recombinant DNA molecules and to host cells
20 transformed therewith. Using standard methodology well known in the art and described briefly below, a recombinant DNA molecule comprising a vector, for example, a Baculovirus transfer vector and a DNA fragment encoding human lactoferrin, for example,
25 HLF 1212 or 1213, can be constructed without undue experimentation.

The methods of choice is the Baculovirus-insect cell expression system (M.D. Summers and G.E. Smith, *Texas Agriculture Experiment Station Bulletin* No. 1555, (1987);
30 V.A. Luckow et al., *Bio/technology* 6:47-55 (1988)). This system has been used successfully to produce commercial quantities of recombinant mammalian glycoproteins. Other expression systems known in

the art can also be used to produce the recombinant protein, for example, yeast, bacterial or mammalian cells.

5 The 2.2 Kb Eco-R1 fragment containing the entire human lactoferrin coding region may be removed from plasmid HLF 1212 or HLF 1213. The lactoferrin cDNA may be subcloned into Baculovirus transfer vector pAc 700 series (T. Maniatis et al., *Molecular Cloning: a laboratory manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, New York).

10 Recombinant plasmid (Achlf) may be co-transfected into Sf9 cells along with wild-type AcNPV viral DNA by calcium phosphate transfection procedure (M.D. Summus and G.E. Smith). In vivo homologous

15 recombination between the polyhedron sequences in the wild type viral DNA and the recombinant plasmid results in the generation of recombinant viruses coding for a fused gene product. The recombinant viruses may be plaque purified by screening for the

20 occlusion negative (polyhedron negative) phenotype or by colony hybridization using ³²P-DNA probes covering the HLF-coding region. Characterization of the recombinant viral DNA may be carried out as described by Maniatis et al. Sf9 cells may be

25 plated in 24-well dishes (Costar) at 3×10^5 cells/well and allowed to attach for 2 hours in complete Grace's medium. Cells are then infected with wild type AcNPV or recombinant virus AchLF. Two days post-infection, the cell layer and the

30 condition medium may be collected and assayed for the presence of hLF. HLF can be analyzed by SDA-PAGE and Western blotting. Iron binding capacity and anti-bacterial activity may also be examined.

The present invention further relates to treatment of antibacterial and antiviral infections using pharmaceutical doses of human lactoferrin of the present invention (HLF 1212 and 1213
5 corresponding to sequence ID Nos. 2 and 4 respectively) or recombinant human lactoferrin protein of the present invention.

The actions of lactoferrin are varied; the best established function is antibacterial (R.R. Arnold et al., *Science* 197:263-265 (1977)). Patients
10 have been found whose neutrophils are deficient in lactoferrin (K.J. Lomax et al., *J. Clin. Invest.* 83:514-519 (1989)). These patients are prone to recurrent infections. Lactoferrin also has been found to
15 decrease release of CSF or monokines, enhancement monocyte natural killer activity, enhancement of hydroxyl radical production and modulate the activation of the complement system (Birgens, *Scand.J. Haematol* 33:225-230 (1984)). There is also early in
20 vivo evidence of lactoferrin antiviral activity.

In the past few years, HIV infection has become a significant health problem. HIV causes morbidity by crippling the body's defense mechanism and allowing development of opportunistic
25 infections. Present treatment is less than ideal and involves treating opportunistic infections as they occur or inhibiting reverse transcriptase. Human lactoferrin is the natural product of the human defense machinery and has been given to
30 patients both orally and intravenously with no side effects. Due to its bacteriocidal, antifungal, and immunoregulatory activity, administering pharmaceutical acceptable doses of lactoferrin of

the present invention could prove an effective agent to treat patients with AIDS or patients with neutropenia.

Other possible uses of the human lactoferrin of the present invention include treatment of lactoferrin in pharmaceutical doses, either orally or intravenously to patients with skin infections (burn patients), gastrointestinal bacterial overgrowth syndromes, vaginal infections, septic shock, and numerous other disorders.

In yet another embodiment, the present invention relates to the genomic human lactoferrin promotor region (sequence ID No: 5). This sequence contains the entire human lactoferrin promotor region fragment including exon 1 of human lactoferrin clone 1212.

The 5' genomic regulatory region of the present invention has the ability to regulate DNA in a tissue specific manner, i.e., it can be on in breast tissue and off in skin. It also can be hormonally regulated, i.e., on in mid-cycle menstrual cycle, off at menses. This regulation ability may be used in several ways. Genes targeted for transgenic mice may use the lactoferrin promotor. Genes to be used in therapy of human disease (gene therapy) may be linked to the lactoferrin promotor and thus the therapeutic gene regulated in a tissue specific or hormonal pattern.

The invention is described in further detail in the following non-limited examples.

EXAMPLES

The following procedures and materials were used throughout the Examples.

Human tissue.

5 150 ml of heparinized blood or 5 ml
heparinized bone marrow was obtained from normal
paid donors after informed consent was obtained.
Informed consent and leukemia cells were obtained
from seven patients with acute leukemia undergoing
10 emergent leukapheresis. The FAB classification of
the patients were: two patients with M2, two
patients with M7, and one patient each with M4, M7,
ANLL not further specified, and T-cell ALL.
Nucleated cells were obtained from 80 ml of blood
15 from normal donors after first incubating cells at
37° C for 30 min. in 1:20 diluted methylcellulose
(30 g/500 ml Hank balanced salt solution (HBSS) to
sediment the red blood cells. The leukocyte-rich
fraction was removed, and centrifuged into a pellet
20 at 500 x g for 10 min. at 4° C. Cells from patients
with leukemia were either used fresh or diluted in
RPMI 1640 containing 20% fetal calf serum and 10%
dimethylsulfoxide (DMSO), then frozen at -70° C
until use. Human leukocyte antigen (HLA) typing,
25 cytogenetic analysis, and bone marrow biopsy results
were available for all but one patient who died
shortly after leukapheresis. All cell lines were
originally obtained from ATCC (Rockville, MD) and
maintained at 37° C, 93% humidity, and 5% CO₂.
30 Breast cancer cell lines and HBL 100 (a cell line
derived from a lactating breast) were maintained and
provided by Dr. J. Dirk Iglehart (Department of

Surgery, Duke University). Cells were grown to confluence and separated from dishes with trypsin 0.05%/EDTA (Gibco), washed, and centrifuged. For all samples, DNA was isolated according to standard methodology (W.M. Strauss in Current Protocols in Molecular Biology. F.A. Ausebel, et al., (eds.), pp. 2.2.1 - 2.2.3 1990. Greene Publishing and Wiley-Interscience, New York.

Isolation of cDNA

A Clontech cDNA library from normal human breast tissue (HL 1037b) was plated in host cells Y1090, filter-lifted and probed with mouse lactoferrin cDNA T267 (B.T. Pentecost and C.T. Teng, (1987)). Positive clones were plaque-purified, and the inserts subcloned into the Eco R1 site of Bluescript II SK+ (Stratagene). The recombinant clones were transformed into XL1 Blue cells (Stratagene). A 2.1 Kb insert (HLF 1212) was isolated and sequenced using the dideoxy nucleotide termination reaction and [³²S]dATP label under contract by Lark sequencing company.

Southern Analysis

Ten µg of DNA was digested at 37° C for three hours with Eco R1, Bam H1, Hind III, Pvu II, Pst I, Msp I, Xba I, Hpa II, Mbo I or Sau 3AI under conditions specified by the manufacturer (BRL). Hpa II and Sau 3AI will not cleave DNA when specific bases within their recognition sites are methylated. Msp I and Mbo I respectively, recognize these same sites and are methylation insensitive. DNA was loaded into 0.7, 0.8, or 1.2% agarose gels, run

overnight, and transferred either to Genescreen Plus (nylon, DuPont) or BA-S NC (nitrocellulose, Schleicher & Schuel). Lactoferrin cDNA was removed from plasmid with Eco RI, redigested with Pst I, and gel purified. Both fragments were labeled with [³²P]dCTP using a random primer kit (Stratagene) to a specific activity of 1×10^9 . Hybridization was performed exactly according to Genescreen instructions or a modification of BA-S NC instructions (hybridization solution - 50% formamide, 5x SSPE, 1% SDS, 4x Denhardt, 100 µg/ml single stranded DNA, 7.5% dextran, pre-hybridization solution - the same as above with 5% formamide and no dextran). Filters were washed at high stringency at 60° C and exposed to Kodak XOMAT AR film using intensifying screens for 3-7 days. DNA from normal and leukemic cells was probed with histone cDNA (Oncore) as a control; no polymorphic pattern was found.

Immunocytochemistry

Antibody against human milk lactoferrin (Sigma) was raised in rabbits and the IgG fraction was prepared as described previously (C.T. Teng et al., *Endocrinology* 124:992-999 (1989)). All cell lines, normal cells, and leukemia patient's cells were examined using this antibody. Ten normal bone marrow specimens were stained to define the specific cell in bone marrow that begins to produce lactoferrin. Cells were smeared onto alcohol-washed, pre-cleaned slides, air dried 1 hour, and fixed in 95% methanol, and 1.7% formalin for 10 min. Slides were next rinsed in dH₂O and either air dried and stored in a moisture proof container at 4° C or

used immediately. Staining procedure was followed directions provided with Vector ABC-AP kit using levamisol as the blocking agent, antibody dilution of 1:1500, and hematoxylin (gill #3) counterstain.
5 Three-hundred cells per sample were scored manually as negative, trace, or positive.

Example 1. Immunocytochemical staining.

As shown in Table 1 and Figure 1A, bone marrow lactoferrin began to appear in the myelocyte stage with almost all cells staining positively by
10 the metamyelocyte stage. None of the leukemia cells from patients or leukemia cell lines contained stainable lactoferrin. Occasional positive granulocytes could be seen in with the leukemic
15 cells from patients. Breast cancer cell lines stained negatively for lactoferrin except for 1.5% trace positive cells in SKB R3 (Figure 1B).

Table 1. Immunocytochemical staining of normal bone marrow using anti-lactoferrin antibody

	Blasts and Promyelocytes	Myelocytes	Metamyelocytes	Bands	Neutrophils
Negative	93% [±] (8.6)	30% (20.4)	12% (7.5)	3% (1.2)	1% (1)
Trace	6% (8.2)	38% (8.3)	40% (10.6)	10% (5.2)	2% (2)
Positive	0.3% (0.4)	32% (19.2)	48% (17)	88% (4.5)	97% (2)

α - values represent the mean of 10 bone marrow samples stained with the standard deviation in parenthesis, >300 cells counted per sample.

Example 2. Library screening, isolation and characterization of HLF 1212 clone.

Thirty human lactoferrin clones were isolated from the breast tissue cDNA library. The longest (HLF 1212) was sequenced completely. This clone is 2117 bp's in length and includes a 17 amino acid (AA) leader sequence (no ATG site) and is 4 AA shy of the 3' terminus (Figure 10). The AA sequence coded for by HLF 1212 has 4 sites that differ from the previously published revised AA sequence derived from the protein (B.F. Anderson et al., (1989)). In the sequence of the present invention, there is one insertion (Arginine (Arg) at AA 22, bp 64-6) and three substitutions (Glutamine (Gln) for Asparagine (Asn) at AA 31, bp 91-3; Isoleucine (Ile) for Leucine (Leu) at AA 55, bp 163-5; and Arg for Lysine (Lys) at AA 218, bp 652-4). The first three of these changes are clustered at the 5' end. Contained within HLF 1212, but not in any of the 10 other partially sequenced isolates, is a deleted cytosine at bp 2097 (AA 699) which caused a frame-shift at the 3' end of the protein. This extra base was confirmed by repeated bi-directional sequencing. The deletion at 2097 is now thought to be either a cloning artifact or a rare species of mRNA.

In addition to cDNA of the present invention, three other authors have published lactoferrin cDNA sequence data (T.A. Rado, et al., (1987); M.J. Powell and J.E. Ogden, *Nucleic Acids Res.*, 18:4013, (1990); M.W. Rey et al., *Nucleic Acids Res.*, 18:5288, (1990)). All of these sequences are different, and a comparison between the AA data derived from the protein and sequence changes derived from the cDNA, are presented in Figure 10. When compared to HLF 1212, all of the sequences

contain an extra cytosine at bp 2097 (AA 699). Powell et al., (1990) isolated a 2.3 kb sequence from breast tissue that, except for the extra cytosine, is identical to our cDNA in the areas of overlap. The isolate of the present invention differs from that of Rado's 3' 1023 base fragment in 4 locations (T.A. Rado et al., (1987)) with one resulting difference in the AA sequence (Gly for Ala at AA 486, bp 1456-8). Two silent mutations and the extra cytosine make up the remainder of the changes. Ray et al have also published a cDNA sequence isolated from human mammary tissue that contains two AA changes (Ile for Thr at AA 147, bp 440-2; and Gly for Cys at AA 421, bp 1261-3) and one silent base difference (M.W. Rey et al., (1990)).

Example 3. Evaluation of restriction fragments using lactoferrin HLF 1212 as probe.

The fragments produced by digestion with Eco RI, Bam HI, Hind III, Pst I, Pvu II, Sau 3AI, or Mbo I, were nearly identical whether the DNA was from normal or malignant cells. The fragment patterns produced by these restriction enzymes in DNA from leukemic and breast cancer cells are shown in Figures 2 and 3. Restriction with Msp I indicated the deletion of a 3.5 Kb band in 3 of 10 leukemic cells (Figure 4), 4 of 7 breast cancer cell lines (Figure 5), and a much fainter hybridization of this band in 2 of 9 normal specimens (Figure 4). An extra 1.3 Kb band also occurred in the breast cancer line MDA 175 (Figure 5, lane 7). There was no relationship between the phenotype or chromosome

analysis of the leukemia patients and the Msp I changes.

Fragments produced by Xba I fell into 4 patterns. All patterns contained 4 unchanged bands (~6.5 kb, ~4.2 kb, ~3.0 kb, and ~2.2 kb). Pattern A occurred in 3 of 9 normal samples and contained a 3.5 Kb band and three light < 2.0 kb bands in addition to the unchanged bands (Figure 6, lanes 1, 2, and 7; Figure 7, lane 1). Pattern B was seen in 6 of 9 normal and 3 of 7 leukemia cells from patients and contained extra 3.5, 5.0, and 6.7 Kb bands along with the three light < 2.0 kb bands and the unchanged bands (Figure 6, lanes 3-6, 8, 9, 10, 13, 14; Figure 7, lane 2). The last patterns were only seen in DNA obtained from malignant tissue. In pattern C, an extra 9.0 Kb band together with the 3.5, 5.0, and 6.6 kb and unchanged bands were observed in three leukemia patient samples (Figure 6 lanes 11, 12 (see insert) and lane 16). Also noted is the absence of the light < 2.0 kb bands. Pattern D contained only the 4 unchanged and the three light < 2.0 kb bands and was present in DNA obtained from all three leukemia and all seven breast cancer cell lines, (Figure 6, lanes 17 - 19, and Figure 7, lanes 3 - 9). There was one patient (M2 leukemia) with a restriction pattern like that of the cell lines (Figure 6, lane 15). There were no chromosomal abnormalities, French-American-British (FAB) categories, or phenotypic types associated with any polymorphic Xba I pattern.

Example 4. Isolation and characterization of the genomic lactoferrin promotor region.

5 A human placental DNA library (Clontech) was plated on LE 392 bacterial cells and screened and probed with the 5' end of HLF 1212 (1.3Kb). Positive clones were cut with SAC 1 and rescreened using a 25 base oligonucleotide (synthesized to match Exon 1 of p1212). All SAC 1 fragments from clone 031A were transformed into Bluescript II KS
10 (stratagene) plasmid. Clone 031A-30 was 2.0 kb and hybridized to Exon 1 oligonucleotide probe. This was sequenced using dideoxynucleotide chain termination and synthesized oligonucleotide primers. Sequence ID NO. 5 shows the sequence of the entire fragment
15 (5' - 3') that includes Exon 1.

* * * *

While the foregoing invention has been described in some detail for purpose of clarity and understanding, it will be clear to one skilled in the art from a reading of this disclosure that
20 various changes in form and detail can be made without departing from the true scope of the invention.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Teng, Christina
Panella, Timothy J.
 - (ii) TITLE OF INVENTION: HUMAN LACTOFERRIN
 - (iii) NUMBER OF SEQUENCES: 5
 - (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: CUSHMAN, DARBY & CUSHMAN
 - (B) STREET: 1615 L. STREET N.W., ELEVENTH FLOOR
 - (C) CITY: WASHINGTON
 - (D) STATE: D.C.
 - (E) COUNTRY: USA
 - (F) ZIP: 20036-5601
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: SCOTT, WATSON T.
 - (B) REGISTRATION NUMBER: 26,581
 - (C) REFERENCE/DOCKET NUMBER: WTS/5683/84482/KIK
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (202) 861-3000
 - (B) TELEFAX: (202) 822-0944
 - (C) TELEX: 6714627 CUSH
- (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2117 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

25

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..2117

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CTT	GTC	TTC	CTC	GTC	CTG	CTG	TTC	CTC	GGG	GCC	CTC	GGA	CTG	TGT	CTG	48
Leu	Val	Phe	Leu	Val	Leu	Leu	Phe	Leu	Gly	Ala	Leu	Gly	Leu	Cys	Leu	
1				5					10					15		
GCT	GGC	CGT	AGG	AGA	AGG	AGT	GTT	CAG	TGG	TGC	GCC	GTA	TCC	CAA	CCC	96
Ala	Gly	Arg	Arg	Arg	Arg	Ser	Val	Gln	Trp	Cys	Ala	Val	Ser	Gln	Pro	
			20					25					30			
GAG	GCC	ACA	AAA	TGC	TTC	CAA	TGG	CAA	AGG	AAT	ATG	AGA	AAA	GTG	CGT	144
Glu	Ala	Thr	Lys	Cys	Phe	Gln	Trp	Gln	Arg	Asn	Met	Arg	Lys	Val	Arg	
		35					40					45				
GGC	CCT	CCT	GTC	AGC	TGC	ATA	AAG	AGA	GAC	TCC	CCC	ATC	CAG	TGT	ATC	192
Gly	Pro	Pro	Val	Ser	Cys	Ile	Lys	Arg	Asp	Ser	Pro	Ile	Gln	Cys	Ile	
	50					55					60					
CAG	GCC	ATT	GCG	GAA	AAC	AGG	GCC	GAT	GCT	GTG	ACC	CTT	GAT	GGT	GGT	240
Gln	Ala	Ile	Ala	Glu	Asn	Arg	Ala	Asp	Ala	Val	Thr	Leu	Asp	Gly	Gly	
65					70				75						80	
TTC	ATA	TAC	GAG	GCA	GGC	CTG	GCC	CCC	TAC	AAA	CTG	CGA	CCT	GTA	GCG	288
Phe	Ile	Tyr	Glu	Ala	Gly	Leu	Ala	Pro	Tyr	Lys	Leu	Arg	Pro	Val	Ala	
				85					90					95		
GCG	GAA	GTC	TAC	GGG	ACC	GAA	AGA	CAG	CCA	CGA	ACT	CAC	TAT	TAT	GCC	336
Ala	Glu	Val	Tyr	Gly	Thr	Glu	Arg	Gln	Pro	Arg	Thr	His	Tyr	Tyr	Ala	
			100					105					110			
GTG	GCT	GTG	GTG	AAG	AAG	GGC	GGC	AGC	TTT	CAG	CTG	AAC	GAA	CTG	CAA	384
Val	Ala	Val	Val	Lys	Lys	Gly	Gly	Ser	Phe	Gln	Leu	Asn	Glu	Leu	Gln	
		115					120					125				
GGT	CTG	AAG	TCC	TGC	CAC	ACA	GGC	CTT	CGC	AGG	ACC	GCT	GGA	TGG	AAT	432
Gly	Leu	Lys	Ser	Cys	His	Thr	Gly	Leu	Arg	Arg	Thr	Ala	Gly	Trp	Asn	
	130					135					140					
GTC	CCT	ATA	GGG	ACA	CTT	CGT	CCA	TTC	TTG	AAT	TGG	ACG	GGT	CCA	CCT	480
Val	Pro	Ile	Gly	Thr	Leu	Arg	Pro	Phe	Leu	Asn	Trp	Thr	Gly	Pro	Pro	
145					150					155					160	

26

GAG Glu	CCC Pro	ATT Ile	GAG Glu	GCA Ala 165	GCT Ala	GTG Val	GCC Ala	AGG Arg	TTC Phe 170	TTC Phe	TCA Ser	GCC Ala	AGC Ser	TGT Cys 175	GTT Val	528
CCC Pro	GGT Gly	GCA Ala	GAT Asp 180	AAA Lys	GGA Gly	CAG Gln	TTC Phe	CCC Pro 185	AAC Asn	CTG Leu	TGT Cys	CGC Arg	CTG Leu 190	TGT Cys	GCG Ala	576
GGG Gly	ACA Thr	GGG Gly 195	GAA Glu	AAC Asn	AAA Lys	TGT Cys	GCC Ala 200	TTC Phe	TCC Ser	TCC Ser	CAG Gln	GAA Glu 205	CCG Pro	TAC Tyr	TTC Phe	624
AGC Ser	TAC Tyr 210	TCT Ser	GGT Gly	GCC Ala	TTC Phe	AAG Lys 215	TGT Cys	CTG Leu	AGA Arg	GAC Asp	GGG Gly 220	GCT Ala	GGA Gly	GAC Asp	GTG Val	672
GCT Ala 225	TTT Phe	ATC Ile	AGA Arg	GAG Glu	AGC Ser 230	ACA Thr	GTG Val	TTT Phe	GAG Glu	GAC Asp 235	CTG Leu	TCA Ser	GAC Asp	GAG Glu	GCT Ala 240	720
GAA Glu	AGG Arg	GAC Asp	GAG Glu	TAT Tyr 245	GAG Glu	TTA Leu	CTC Leu	TGC Cys	CCA Pro 250	GAC Asp	AAC Asn	ACT Thr	CGG Arg	AAG Lys 255	CCA Pro	768
GTG Val	GAC Asp	AAG Lys	TTC Phe 260	AAA Lys	GAC Asp	TGC Cys	CAT His	CTG Leu 265	GCC Ala	CGG Arg	GTC Val	CCT Pro	TCT Ser 270	CAT His	GCC Ala	816
GTT Val	GTG Val	GCA Ala 275	CGA Arg	AGT Ser	GTG Val	AAT Asn	GGC Gly 280	AAG Lys	GAG Glu	GAT Asp	GCC Ala	ATC Ile 285	TGG Trp	AAT Asn	CTT Leu	864
CTC Leu 290	CGC Arg	CAG Gln	GCA Ala	CAG Gln	GAA Glu	AAG Lys 295	TTT Phe	GGA Gly	AAG Lys	GAC Asp	AAG Lys 300	TCA Ser	CCG Pro	AAA Lys	TTC Phe	912
CAG Gln 305	CTC Leu	TTT Phe	GGC Gly	TCC Ser	CCT Pro 310	AGT Ser	GGG Gly	CAG Gln	AAA Lys	GAT Asp 315	CTG Leu	CTG Leu	TTC Phe	AAG Lys	GAC Asp 320	960
TCT Ser	GCC Ala	ATT Ile	GGG Gly	TTT Phe 325	TCG Ser	AGG Arg	GTG Val	CCC Pro	CCG Pro 330	AGG Arg	ATA Ile	GAT Asp	TCT Ser 335	GGG Gly 335	CTG Leu	1008
TAC Tyr	CTT Leu	GGC Gly	TCC Ser 340	GGC Gly	TAC Tyr	TTC Phe	ACT Thr	GCC Ala 345	ATC Ile	CAG Gln	AAC Asn	TTG Leu	AGG Arg 350	AAA Lys	AGT Ser	1056

27

GAG Glu	GAG Glu	GAA Glu 355	GTG Val	GCT Ala	GCC Ala	CGG Arg	CGT Arg 360	GCG Ala	CGG Arg	GTC Val	GTG Val	TGG Trp 365	TGT Cys	GCG Ala	GTG Val	1104
GGC Gly 370	GAG Glu	CAG Gln	GAG Glu	CTG Leu	CGC Arg	AAG Lys 375	TGT Cys	AAC Asn	CAG Gln	TGG Trp	AGT Ser 380	GGC Gly	TTG Leu	AGC Ser	GAA Glu	1152
GGC Gly 385	AGC Ser	GTG Val	ACC Thr	TGC Cys	TCC Ser 390	TCG Ser	GCC Ala	TCC Ser	ACC Thr	ACA Thr 395	GAG Glu	GAC Asp	TGC Cys	ATC Ile	GCC Ala 400	1200
CTG Leu	GTG Val	CTG Leu	AAA Lys 405	GGA Gly	GAA Glu	GCT Ala	GAT Asp	GCC Ala	ATG Met 410	AGT Ser	TTG Leu	GAT Asp	GGA Gly	GGA Gly 415	TAT Tyr	1248
GTG Val	TAC Tyr	ACT Thr	GCA Ala 420	GGC Gly	AAA Lys	TGT Cys	GGT Gly	TTG Leu 425	GTG Val	CCT Pro	GTC Val	CTG Leu	GCA Ala 430	GAG Glu	AAC Asn	1296
TAC Tyr	AAA Lys	TCC Ser	CAA Gln 435	CAA Gln	AGC Ser	AGT Ser	GAC Asp 440	CCT Pro	GAT Asp	CCT Pro	AAC Asn	TGT Cys 445	GTG Val	GAT Asp	AGA Arg	1344
CCT Pro 450	GTG Val	GAA Glu	GGA Gly	TAT Tyr	CTT Leu	GCT Ala 455	GTG Val	GCG Ala	GTG Val	GTT Val	AGG Arg 460	AGA Arg	TCA Ser	GAC Asp	ACT Thr	1392
AGC Ser 465	CTT Leu	ACC Thr	TGG Trp	AAC Asn	TCT Ser 470	GTG Val	AAA Lys	GGC Gly	AAG Lys	AAG Lys 475	TCC Ser	TGC Cys	CAC His	ACC Thr	GCC Ala 480	1440
GTG Val	GAC Asp	AGG Arg	ACT Thr	GCA Ala 485	GGC Gly	TGG Trp	AAT Asn	ATC Ile	CCC Pro 490	ATG Met	GGC Gly	CTG Leu	CTC Leu	TTC Phe 495	AAC Asn	1488
CAG Gln	ACG Thr	GGC Gly	TCC Ser 500	TGC Cys	AAA Lys	TTT Phe	GAT Asp	GAA Glu	TAT Tyr	TTC Phe	AGT Ser	CAA Gln	AGC Ser	TGT Cys	GCC Ala	1536
CCT Pro	GGG Gly	TCT Ser 515	GAC Asp	CCG Pro	AGA Arg	TCT Ser	AAT Asn 520	CTC Leu	TGT Cys	GCT Ala	CTG Leu	TGT Cys 525	ATT Ile	GGC Gly	GAC Asp	1584
GAG Glu 530	CAG Gln	GGT Gly	GAG Glu	AAT Asn	AAG Lys	TGC Cys 535	GTG Val	CCC Pro	AAC Asn	AGC Ser	AAC Asn 540	GAG Glu	AGA Arg	TAC Tyr	TAC Tyr	1632

[illegible]

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 705 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Ala Gly Arg Arg Arg Arg Ser Val Gln Trp Cys Ala Val Ser Gln Pro
          20             25             30
Glu Ala Thr Lys Cys Phe Gln Trp Gln Arg Asn Met Arg Lys Val Arg
          35             40             45
Gly Pro Pro Val Ser Cys Ile Lys Arg Asp Ser Pro Ile Gln Cys Ile
          50             55             60
Gln Ala Ile Ala Glu Asn Arg Ala Asp Ala Val Thr Leu Asp Gly Gly
 65             70             75             80
Phe Ile Tyr Glu Ala Gly Leu Ala Pro Tyr Lys Leu Arg Pro Val Ala
          85             90             95
Ala Glu Val Tyr Gly Thr Glu Arg Gln Pro Arg Thr His Tyr Tyr Ala
          100            105            110
Val Ala Val Val Lys Lys Gly Gly Ser Phe Gln Leu Asn Glu Leu Gln
          115            120            125
Gly Leu Lys Ser Cys His Thr Gly Leu Arg Arg Thr Ala Gly Trp Asn
          130            135            140
Val Pro Ile Gly Thr Leu Arg Pro Phe Leu Asn Trp Thr Gly Pro Pro
          145            150            155            160
Glu Pro Ile Glu Ala Ala Val Ala Arg Phe Phe Ser Ala Ser Cys Val
          165            170            175
Pro Gly Ala Asp Lys Gly Gln Phe Pro Asn Leu Cys Arg Leu Cys Ala
          180            185            190
Gly Thr Gly Glu Asn Lys Cys Ala Phe Ser Ser Gln Glu Pro Tyr Phe
          195            200            205

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30

Ser Tyr Ser Gly Ala Phe Lys Cys Leu Arg Asp Gly Ala Gly Asp Val
 210 215 220
 Ala Phe Ile Arg Glu Ser Thr Val Phe Glu Asp Leu Ser Asp Glu Ala
 225 230 235 240
 Glu Arg Asp Glu Tyr Glu Leu Leu Cys Pro Asp Asn Thr Arg Lys Pro
 245 250 255
 Val Asp Lys Phe Lys Asp Cys His Leu Ala Arg Val Pro Ser His Ala
 260 265 270
 Val Val Ala Arg Ser Val Asn Gly Lys Glu Asp Ala Ile Trp Asn Leu
 275 280 285
 Leu Arg Gln Ala Gln Glu Lys Phe Gly Lys Asp Lys Ser Pro Lys Phe
 290 295 300
 Gln Leu Phe Gly Ser Pro Ser Gly Gln Lys Asp Leu Leu Phe Lys Asp
 305 310 315 320
 Ser Ala Ile Gly Phe Ser Arg Val Pro Pro Arg Ile Asp Ser Gly Leu
 325 330 335
 Tyr Leu Gly Ser Gly Tyr Phe Thr Ala Ile Gln Asn Leu Arg Lys Ser
 340 345 350
 Glu Glu Glu Val Ala Ala Arg Arg Ala Arg Val Val Trp Cys Ala Val
 355 360 365
 Gly Glu Gln Glu Leu Arg Lys Cys Asn Gln Trp Ser Gly Leu Ser Glu
 370 375 380
 Gly Ser Val Thr Cys Ser Ser Ala Ser Thr Thr Glu Asp Cys Ile Ala
 385 390 395 400
 Leu Val Leu Lys Gly Glu Ala Asp Ala Met Ser Leu Asp Gly Gly Tyr
 405 410 415
 Val Tyr Thr Ala Gly Lys Cys Gly Leu Val Pro Val Leu Ala Glu Asn
 420 425 430
 Tyr Lys Ser Gln Gln Ser Ser Asp Pro Asp Pro Asn Cys Val Asp Arg
 435 440 445
 Pro Val Glu Gly Tyr Leu Ala Val Ala Val Val Arg Arg Ser Asp Thr
 450 455 460
 Ser Leu Thr Trp Asn Ser Val Lys Gly Lys Lys Ser Cys His Thr Ala
 465 470 475 480

Val Asp Arg Thr Ala Gly Trp Asn Ile Pro Met Gly Leu Leu Phe Asn
 485 490 495
 Gln Thr Gly Ser Cys Lys Phe Asp Glu Tyr Phe Ser Gln Ser Cys Ala
 500 505 510
 Pro Gly Ser Asp Pro Arg Ser Asn Leu Cys Ala Leu Cys Ile Gly Asp
 515 520 525
 Glu Gln Gly Glu Asn Lys Cys Val Pro Asn Ser Asn Glu Arg Tyr Tyr
 530 535 540
 Gly Tyr Thr Gly Ala Phe Arg Cys Leu Ala Glu Asn Ala Gly Asp Val
 545 550 555 560
 Ala Phe Val Lys Asp Val Thr Val Leu Gln Asn Thr Asp Gly Asn Asn
 565 570 575
 Asn Glu Ala Trp Ala Lys Asp Leu Lys Leu Ala Asp Phe Ala Leu Leu
 580 585 590
 Cys Leu Asp Gly Lys Arg Lys Pro Val Thr Glu Ala Arg Ser Cys His
 595 600 605
 Leu Ala Met Ala Pro Asn His Ala Val Val Ser Arg Met Asp Lys Val
 610 615 620
 Glu Arg Leu Lys Gln Val Leu Leu His Gln Gln Ala Lys Phe Gly Arg
 625 630 635 640
 Asn Gly Ser Asp Cys Pro Asp Lys Phe Cys Leu Phe Gln Ser Glu Thr
 645 650 655
 Lys Asn Leu Leu Phe Asn Asp Asn Thr Glu Cys Leu Ala Arg Leu His
 660 665 670
 Gly Lys Thr Thr Tyr Glu Lys Tyr Leu Gly Pro Gln Tyr Val Ala Gly
 675 680 685
 Ile Thr Asn Leu Lys Lys Cys Ser Thr Ser Pro Ser Trp Lys Pro Val
 690 695 700
 Asn
 705

32

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2124 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..2124

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATG AAA CTT GTC TTC CTC GTC CTG CTG TTC CTC GGG GCC CTC GGA CTG	48
Met Lys Leu Val Phe Leu Val Leu Leu Phe Leu Gly Ala Leu Gly Leu	
1 5 10 15	
TGT CTG GCT GGC CGT AGG AGA AGG AGT GTT CAG TGG TGC GCC GTA TCC	96
Cys Leu Ala Gly Arg Arg Arg Arg Ser Val Gln Trp Cys Ala Val Ser	
20 25 30	
CAA CCC GAG GCC ACA AAA TGC TTC CAA TGG CAA AGG AAT ATG AGA AAA	144
Gln Pro Glu Ala Thr Lys Cys Phe Gln Trp Gln Arg Asn Met Arg Lys	
35 40 45	
GTG CGT GGC CCT CCT GTC AGC TGC ATA AAG AGA GAC TCC CCC ATC CAG	192
Val Arg Gly Pro Pro Val Ser Cys Ile Lys Arg Asp Ser Pro Ile Gln	
50 55 60	
TGT ATC CAG GCC ATT GCG GAA AAC AGG GCC GAT GCT GTG ACC CTT GAT	240
Cys Ile Gln Ala Ile Ala Glu Asn Arg Ala Asp Ala Val Thr Leu Asp	
65 70 75 80	
GGT GGT TTC ATA TAC GAG GCA GGC CTG GCC CCC TAC AAA CTG CGA CCT	288
Gly Gly Phe Ile Tyr Glu Ala Gly Leu Ala Pro Tyr Lys Leu Arg Pro	
85 90 95	
GTA GCG GCG GAA GTC TAC GGG ACC GAA AGA CAG CCA CGA ACT CAC TAT	336
Val Ala Ala Glu Val Tyr Gly Thr Glu Arg Gln Pro Arg Thr His Tyr	
100 105 110	

TAT GCC GTG GCT GTG GTG AAG AAG GGC GGC AGC TTT CAG CTG AAC GAA	384
Tyr Ala Val Ala Val Val Lys Lys Gly Gly Ser Phe Gln Leu Asn Glu	
115 120 125	
CTG CAA GGT CTG AAG TCC TGC CAC ACA GGC CTT CGC AGG ACC GCT GGA	432
Leu Gln Gly Leu Lys Ser Cys His Thr Gly Leu Arg Arg Thr Ala Gly	
130 135 140	
TGG AAT GTC CCT ATA GGG ACA CTT CGT CCA TTC TTG AAT TGG ACG GGT	480
Trp Asn Val Pro Ile Gly Thr Leu Arg Pro Phe Leu Asn Trp Thr Gly	
145 150 155 160	
CCA CCT GAG CCC ATT GAG GCA GCT GTG GCC AGG TTC TTC TCA GCC AGC	528
Pro Pro Glu Pro Ile Glu Ala Ala Val Ala Arg Phe Phe Ser Ala Ser	
165 170 175	
TGT GTT CCC GGT GCA GAT AAA GGA CAG TTC CCC AAC CTG TGT CGC CTG	576
Cys Val Pro Gly Ala Asp Lys Gly Gln Phe Pro Asn Leu Cys Arg Leu	
180 185 190	
TGT GCG GGG ACA GGG GAA AAC AAA TGT GCC TTC TCC TCC CAG GAA CCG	624
Cys Ala Gly Thr Gly Glu Asn Lys Cys Ala Phe Ser Ser Gln Glu Pro	
195 200 205	
TAC TTC AGC TAC TCT GGT GCC TTC AAG TGT CTG AGA GAC GGG GCT GGA	672
Tyr Phe Ser Tyr Ser Gly Ala Phe Lys Cys Leu Arg Asp Gly Ala Gly	
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GAC GTG GCT TTT ATC AGA GAG AGC ACA GTG TTT GAG GAC CTG TCA GAC	720
Asp Val Ala Phe Ile Arg Glu Ser Thr Val Phe Glu Asp Leu Ser Asp	
225 230 235 240	
GAG GCT GAA AGG GAC GAG TAT GAG TTA CTC TGC CCA GAC AAC ACT CGG	768
Glu Ala Glu Arg Asp Glu Tyr Glu Leu Leu Cys Pro Asp Asn Thr Arg	
245 250 255	
AAG CCA GTG GAC AAG TTC AAA GAC TGC CAT CTG GCC CGG GTC CCT TCT	816
Lys Pro Val Asp Lys Phe Lys Asp Cys His Leu Ala Arg Val Pro Ser	
260 265 270	

CAT GCC GTT GTG GCA CGA AGT GTG AAT GGC AAG GAG GAT GCC ATC TGG	864
His Ala Val Val Ala Arg Ser Val Asn Gly Lys Glu Asp Ala Ile Trp	
275 280 285	
AAT CTT CTC CGC CAG GCA CAG GAA AAG TTT GGA AAG GAC AAG TCA CCG	912
Asn Leu Leu Arg Gln Ala Gln Glu Lys Phe Gly Lys Asp Lys Ser Pro	
290 295 300	
AAA TTC CAG CTC TTT GGC TCC CCT AGT GGG CAG AAA GAT CTG CTG TTC	960
Lys Phe Gln Leu Phe Gly Ser Pro Ser Gly Gln Lys Asp Leu Leu Phe	
305 310 315 320	
AAG GAC TCT GCC ATT GGG TTT TCG AGG GTG CCC CCG AGG ATA GAT TCT	1008
Lys Asp Ser Ala Ile Gly Phe Ser Arg Val Pro Pro Arg Ile Asp Ser	
325 330 335	
GGG CTG TAC CTT GGC TCC GGC TAC TTC ACT GCC ATC CAG AAC TTG AGG	1056
Gly Leu Tyr Leu Gly Ser Gly Tyr Phe Thr Ala Ile Gln Asn Leu Arg	
340 345 350	
AAA AGT GAG GAG GAA GTG GCT GCC CGG CGT GCG CGG GTC GTG TGG TGT	1104
Lys Ser Glu Glu Glu Val Ala Ala Arg Arg Ala Arg Val Val Trp Cys	
355 360 365	
GCG GTG GGC GAG CAG GAG CTG CGC AAG TGT AAC CAG TGG AGT GGC TTG	1152
Ala Val Gly Glu Gln Glu Leu Arg Lys Cys Asn Gln Trp Ser Gly Leu	
370 375 380	
AGC GAA GGC AGC GTG ACC TGC TCC TCG GCC TCC ACC ACA GAG GAC TGC	1200
Ser Glu Gly Ser Val Thr Cys Ser Ser Ala Ser Thr Thr Glu Asp Cys	
385 390 395 400	
ATC GCC CTG GTG CTG AAA GGA GAA GCT GAT GCC ATG AGT TTG GAT GGA	1248
Ile Ala Leu Val Leu Lys Gly Glu Ala Asp Ala Met Ser Leu Asp Gly	
405 410 415	
GGA TAT GTG TAC ACT GCA GGC AAA TGT GGT TTG GTG CCT GTC CTG GCA	1296
Gly Tyr Val Tyr Thr Ala Gly Lys Cys Gly Leu Val Pro Val Leu Ala	
420 425 430	

35

GAG AAC TAC AAA TCC CAA CAA AGC AGT GAC CCT GAT CCT AAC TGT GTG	1344
Glu Asn Tyr Lys Ser Gln Gln Ser Ser Asp Pro Asp Pro Asn Cys Val	
435 440 445	
GAT AGA CCT GTG GAA GGA TAT CTT GCT GTG GCG GTG GTT AGG AGA TCA	1392
Asp Arg Pro Val Glu Gly Tyr Leu Ala Val Ala Val Val Arg Arg Ser	
450 455 460	
GAC ACT AGC CTT ACC TGG AAC TCT GTG AAA GGC AAG AAG TCC TGC CAC	1440
Asp Thr Ser Leu Thr Trp Asn Ser Val Lys Gly Lys Lys Ser Cys His	
465 470 475 480	
ACC GCC GTG GAC AGG ACT GCA GGC TGG AAT ATC CCC ATG GGC CTG CTC	1488
Thr Ala Val Asp Arg Thr Ala Gly Trp Asn Ile Pro Met Gly Leu Leu	
485 490 495	
TTC AAC CAG ACG GGC TCC TGC AAA TTT GAT GAA TAT TTC AGT CAA AGC	1536
Phe Asn Gln Thr Gly Ser Cys Lys Phe Asp Glu Tyr Phe Ser Gln Ser	
500 505 510	
TGT GCC CCT GGG TCT GAC CCG AGA TCT AAT CTC TGT GCT CTG TGT ATT	1584
Cys Ala Pro Gly Ser Asp Pro Arg Ser Asn Leu Cys Ala Leu Cys Ile	
515 520 525	
GGC GAC GAG CAG GGT GAG AAT AAG TGC GTG CCC AAC AGC AAC GAG AGA	1632
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530 535 540	
TAC TAC GGC TAC ACT GGG GCT TTC CGG TGC CTG GCT GAG AAT GCT GGA	1680
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545 550 555 560	
GAC GTT GCA TTT GTG AAA GAT GTC ACT GTC TTG CAG AAC ACT GAT GGA	1728
Asp Val Ala Phe Val Lys Asp Val Thr Val Leu Gln Asn Thr Asp Gly	
565 570 575	
AAT AAC AAT GAG GCA TGG GCT AAG GAT TTG AAG CTG GCA GAC TTT GCG	1776
Asn Asn Asn Glu Ala Trp Ala Lys Asp Leu Lys Leu Ala Asp Phe Ala	
580 585 590	

CTG CTG TGC CTC GAT GGC AAA CGG AAG CCT GTG ACT GAG GCT AGA AGC	1824
Leu Leu Cys Leu Asp Gly Lys Arg Lys Pro Val Thr Glu Ala Arg Ser 595 600 605	
TGC CAT CTT GCC ATG GCC CCG AAT CAT GCC GTG GTG TCT CGG ATG GAT	1872
Cys His Leu Ala Met Ala Pro Asn His Ala Val Val Ser Arg Met Asp 610 615 620	
AAG GTG GAA CGC CTG AAA CAG GTG TTG CTC CAC CAA CAG GCT AAA TTT	1920
Lys Val Glu Arg Leu Lys Gln Val Leu Leu His Gln Gln Ala Lys Phe 625 630 635 640	
GGG AGA AAT GGA TCT GAC TGC CCG GAC AAG TTT TGC TTA TTC CAG TCT	1968
Gly Arg Asn Gly Ser Asp Cys Pro Asp Lys Phe Cys Leu Phe Gln Ser 645 650 655	
GAA ACC AAA AAC CTT CTG TTC AAT GAC AAC ACT GAG TGT CTG GCC AGA	2016
Glu Thr Lys Asn Leu Leu Phe Asn Asp Asn Thr Glu Cys Leu Ala Arg 660 665 670	
CTC CAT GGC AAA ACA ACA TAT GAA AAA TAT TTG GGA CCA CAG TAT GTC	2064
Leu His Gly Lys Thr Thr Tyr Glu Lys Tyr Leu Gly Pro Gln Tyr Val 675 680 685	
GCA GGC ATT ACT AAT CTG AAA AAG TGC TCA ACC TCC CCC CTC CTG GAA	2112
Ala Gly Ile Thr Asn Leu Lys Lys Cys Ser Thr Ser Pro Leu Leu Glu 690 695 700	
GCC TGT GAA TTC	2124
Ala Cys Glu Phe 705	

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(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 708 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

```

Met Lys Leu Val Phe Leu Val Leu Leu Phe Leu Gly Ala Leu Gly Leu
 1             5             10             15
Cys Leu Ala Gly Arg Arg Arg Arg Ser Val Gln Trp Cys Ala Val Ser
          20             25             30
Gln Pro Glu Ala Thr Lys Cys Phe Gln Trp Gln Arg Asn Met Arg Lys
      35             40             45
Val Arg Gly Pro Pro Val Ser Cys Ile Lys Arg Asp Ser Pro Ile Gln
      50             55             60
Cys Ile Gln Ala Ile Ala Glu Asn Arg Ala Asp Ala Val Thr Leu Asp
      65             70             75             80
Gly Gly Phe Ile Tyr Glu Ala Gly Leu Ala Pro Tyr Lys Leu Arg Pro
          85             90             95
Val Ala Ala Glu Val Tyr Gly Thr Glu Arg Gln Pro Arg Thr His Tyr
          100             105             110
Tyr Ala Val Ala Val Val Lys Lys Gly Gly Ser Phe Gln Leu Asn Glu
          115             120             125
Leu Gln Gly Leu Lys Ser Cys His Thr Gly Leu Arg Arg Thr Ala Gly
          130             135             140
Trp Asn Val Pro Ile Gly Thr Leu Arg Pro Phe Leu Asn Trp Thr Gly
          145             150             155             160
Pro Pro Glu Pro Ile Glu Ala Ala Val Ala Arg Phe Phe Ser Ala Ser
          165             170             175
Cys Val Pro Gly Ala Asp Lys Gly Gln Phe Pro Asn Leu Cys Arg Leu
          180             185             190
Cys Ala Gly Thr Gly Glu Asn Lys Cys Ala Phe Ser Ser Gln Glu Pro
          195             200             205

```

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Tyr Phe Ser Tyr Ser Gly Ala Phe Lys Cys Leu Arg Asp Gly Ala Gly
 210 215 220
 Asp Val Ala Phe Ile Arg Glu Ser Thr Val Phe Glu Asp Leu Ser Asp
 225 230 235 240
 Glu Ala Glu Arg Asp Glu Tyr Glu Leu Leu Cys Pro Asp Asn Thr Arg
 245 250 255
 Lys Pro Val Asp Lys Phe Lys Asp Cys His Leu Ala Arg Val Pro Ser
 260 265 270
 His Ala Val Val Ala Arg Ser Val Asn Gly Lys Glu Asp Ala Ile Trp
 275 280 285
 Asn Leu Leu Arg Gln Ala Gln Glu Lys Phe Gly Lys Asp Lys Ser Pro
 290 295 300
 Lys Phe Gln Leu Phe Gly Ser Pro Ser Gly Gln Lys Asp Leu Leu Phe
 305 310 315 320
 Lys Asp Ser Ala Ile Gly Phe Ser Arg Val Pro Pro Arg Ile Asp Ser
 325 330 335
 Gly Leu Tyr Leu Gly Ser Gly Tyr Phe Thr Ala Ile Gln Asn Leu Arg
 340 345 350
 Lys Ser Glu Glu Glu Val Ala Ala Arg Arg Ala Arg Val Val Trp Cys
 355 360 365
 Ala Val Gly Glu Gln Glu Leu Arg Lys Cys Asn Gln Trp Ser Gly Leu
 370 375 380
 Ser Glu Gly Ser Val Thr Cys Ser Ser Ala Ser Thr Thr Glu Asp Cys
 385 390 395 400
 Ile Ala Leu Val Leu Lys Gly Glu Ala Asp Ala Met Ser Leu Asp Gly
 405 410 415
 Gly Tyr Val Tyr Thr Ala Gly Lys Cys Gly Leu Val Pro Val Leu Ala
 420 425 430
 Glu Asn Tyr Lys Ser Gln Gln Ser Ser Asp Pro Asp Pro Asn Cys Val
 435 440 445
 Asp Arg Pro Val Glu Gly Tyr Leu Ala Val Ala Val Val Arg Arg Ser
 450 455 460
 Asp Thr Ser Leu Thr Trp Asn Ser Val Lys Gly Lys Lys Ser Cys His
 465 470 475 480

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2086 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CGAGGATCAT GGCTCACTGC CACCTTCATC TCCCAGGCTC AAATGGTCCT CCCACTTTAG	60
CCTCCCAAGT AGCTGGGACC ATAGGCATAC ACCACCATGC TGGGCTAATT TTTGTATTTT	120
TTGTAGAGAT GGGGGTTTCC CTATGAAGCC CAGGCTAGTC TTGAACTCCT GGGCTCAAGC	180
GATCCTCCCA TCTTGGCCTC CCAAAGTGCT GGGATTACAG GCATGAGCCA CTGTGCCCTG	240
CCTAGTTACT CTTGGGCTAA GTTCACATCC ATACACACAG GATATTCTTT CTGAGGCCCC	300
CAATGTGTCC CACAGGCACC ATGCTGTATG TGACACTCCC CTAGAGATGG ATGTTTAGTT	360
TGCTTCCAAC TGATTAATGG CATGCAGTGG TGCCTGGAAA CATTTGTACC TGGGGTGCTG	420
TGTGTCATGG GAATGTATTT ACGAGATGTA TTCTTAGAAG CAGTATTCTA GCTTTTGAAT	480
TTTAAAATCT GACATTTATG GCGATTGTTA AAATGAGGTT ACCATTCCT ACTGAATACT	540
ATCAACACCA AAAAAGAAGA AGGAGGAGAT GGAGAAAAA AAGACAAAA AAAAAAAGT	600
GGTAGGGCAT CTTAGCCATA GGGCATCTTT CTCATTGGCA AATAAGAACA TGGAACCAGC	660
CTTGGGTGGT GGCCATTCCC CTCTGAGGTC CCTGTCTGTT TTCTGGGAGC TGTATTGTGG	720
GTCTCAGCAG GGCAGGGAGA TACCCCATGG GCAGCTTGCC TGAGACTCTG GGCAGCCTCT	780
CTTTTCTCTG TCAGCTGTCC CTAGGCTGCT GCTGGGGGTG GTCGGGTCAT CTTTTCAACT	840
CTCAGCTCAC TGCTGAGCCA AGGTGAAAGC AAACCCACCT GCCCTAACTG GCTCCTAGGC	900
ACCTTCAAGG TCATCTGCTG AAGAAGATAG CAGTCTCACA GGTCAAGGCG ATCTTCAAGT	960
AAAGACCCTC TGCTCTGTGT CCTGCCCTCT AGAAGGCACT GAGACCAGAG CTGGGACAGG	1020
GCTCAGGGGG CTGCGACTCC TAGGGGCTTG CAGACCTAGT GGGAGAGAAA GAACATCGCA	1080

GCAGCCAGGC AGAACCAGGA CAGGTGAGGT GCAGGCTGGC TTCCTCTCG CAGCGCGGTG 1140
TGGAGTCCTG TCCTGCCTCA GGGCTTTTCG GAGCCTGGAT CCTCAAGGAA CAAGTAGACC 1200
TGGCCGCGGG GAGTGGGGAG GGAAGGGGTG TCTATTGGGC AACAGGGCGG GGCAAAGCCC 1260
TGAATAAAGG GGCGCAGGGC AGGCGCAAGT GGCAGAGCCT TCGTTTGCCA AGTCGCCTCC 1320
AGACCGCAGA CATGAAACTT GTCTTCCTCG TCCTGCTGTT CCTCGGGGCC CTCGGTGAGT 1380
GCAGGTGCCT GGGGGCGCGA GCCGCCTGAT GGGCGTCTCC TGCGCCCTGT CTGCTAGGCG 1440
CTTTGGTCCC TGTGTCCGGT TGGCTGGGCG CGGGGTCTCT GCGCCCCGCG GTCCCAGCGC 1500
CTACAGCCGG GAGGCGGCCC GGACGCGGGG CCAGTCTCTT TCCCACATGG GGAGGAACAG 1560
GAGCTGGGCT CCTCAAGCCG GATCGGGGCA CGCCTAGCTC TGCTCAGAGC TTCTCAAAAG 1620
GCCTCCCAGG CCCCTGTCCC TTTGTGTCCC GCCTAAGGAT TTGGTCCCCA TTGTATTGTG 1680
ACATGCGTTT TACCTGGGAG GAAAGTGAGG CTCAGAGAGG GTGAGCGACT AGCTCAAGGA 1740
CCCTAGTCCA GATCCTAGCT CCTGCGAGGA CTGTGAGACC CCAGCAAGAC CGAGCCTTTA 1800
TGAGACTTAG TTTCTTCACT TAAAGAAACG GCCTAACCAT GGGTCCACAG GGTGTGTGAGG 1860
AGGAGATGGG GCATTCGCAC ACCTTCCGTG GCAGAGGGTT GTGGAGGGGT GCGGTGCTCC 1920
TGATGGAACC CTGTGTCAGA GGGTTTGAGA GGGAAATGTC AGCCAAACAG AAGGAAGGAG 1980
CAGAAGGAAG GAAACAATTG TCAGTTCCAT AACCAAAGTA ATTTCTCGGG TGCTCAGAGG 2040
GCACTCCCCA GCGCTGCACA TTAGTGACCT AAATGCGTGA GTGCGG 2086

WHAT IS CLAIMED IS:

1. A DNA segment encoding human lactoferrin according to sequence I.D. No.: 1.
- 5 2. Human lactoferrin protein according to sequence I.D. No.: 2.
3. A DNA promotor region for human lactoferrin according to sequence I.D. No.: 5 and allelic variations thereof.
- 10 4. A recombinant DNA construct comprising:
 - i) said DNA segment according to claim 1 and
 - ii) a vector
- 15 5. The DNA construct according to claim 4 further comprising the regulating sequence according to sequence I.D. No.: 5 or portion thereof operatively linked to said DNA fragment.
6. The DNA construct according to claim 4 or 5 wherein said vector is pAc 700 series.
7. A host cell comprising said DNA construct according to claim 4 or 5.
- 20 8. The cell according to claim 7, wherein said host cell is Sf9 cells.
9. A recombinant lactoferrin protein expressed in the host cell of claim 7.
- 25 10. A method of treating a condition in a patient characterization by a deficiency in

lactoferrin, administering to said patient an amount of human lactoferrin according to claims 2 or 9 sufficient to eliminate said deficiency.

- 5 11. The method of claim 10 wherein said condition is neutropenia, AIDS, skin infection, gastrointestinal bacterial overgrowth syndrome, vaginal infection or septic shock.
- 10 12. A method of diagnosing malignancy in a biological sample comprising the steps of:
- i) isolating DNA from said biological sample and normal control sample
 - ii) cutting said DNA with restriction enzyme, Xba I,
 - 15 iii) hybridizing said cut DNA of step (ii) with a DNA segment according to claim 1 or 2 or portion thereof under conditions such that hybridization is effected and
 - 20 iv) comparing the hybridization products of step 3 from said biological sample and normal sample to each other.
- 25 13. A method of detecting recovery of a disease in a patient given a therapeutic comprising the steps of:
- i) isolating DNA from a biological sample of said patient and normal human control sample,
 - ii) cutting said DNA with Xba I,
 - 30 iii) hybridizing said cut DNA of step (ii) with a DNA segment according to claim 1 or

portion thereof under conditions such that hybridization is effected and

- iv) comparing the hybridization products of the biological sample in step 3 to the hybridization products of normal sample in step 3 to determine the relatedness to normal samples.

14. A method for detecting insertions, deletions or mutations surrounding the human lactoferrin gene comprising the steps of

- i) isolating DNA from a biological sample suspected of having said insertion, deletion or mutation,
- ii) amplifying said DNA using the DNA fragment of claim 1 or portion thereof in a polymerase chain reaction,
- iii) cutting said amplified DNA with restriction enzyme Xbu I,
- iv) hybridizing said DNA from step (iii) with the DNA fragment according to claim 1 or portion thereof under conditions such that hybridization is effected and
- v) sequencing said DNA of step (iv).

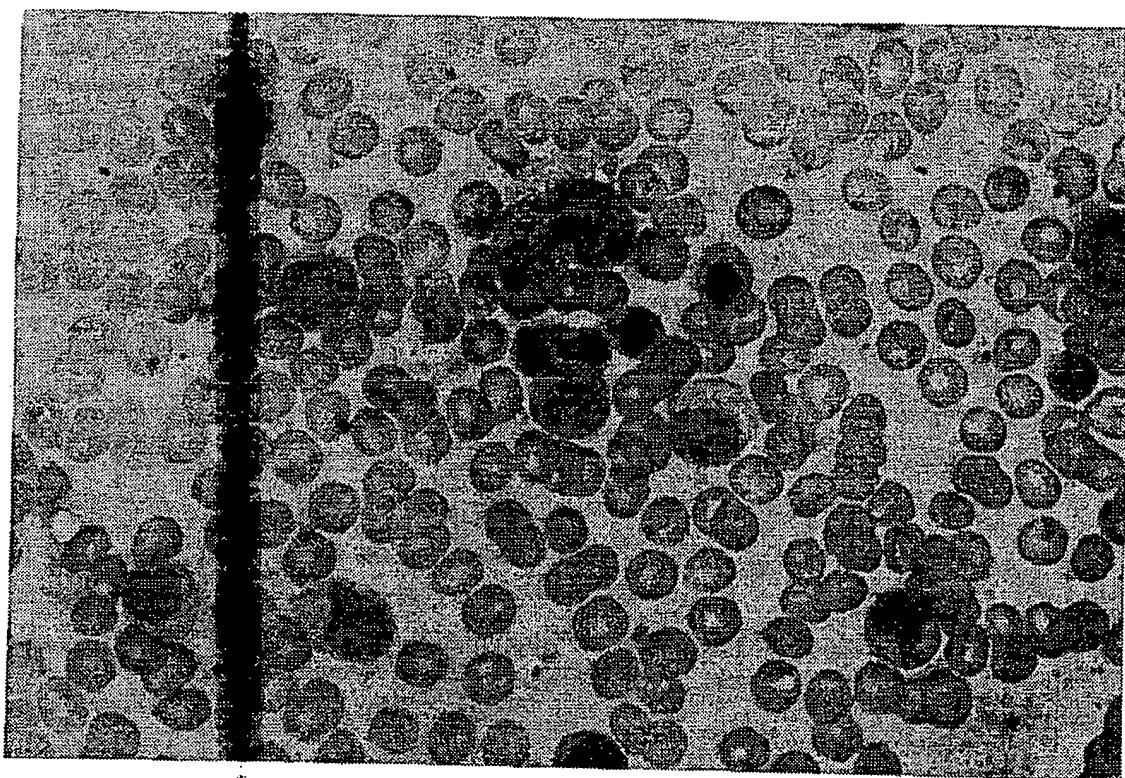


FIG. 1A



FIG. 1B

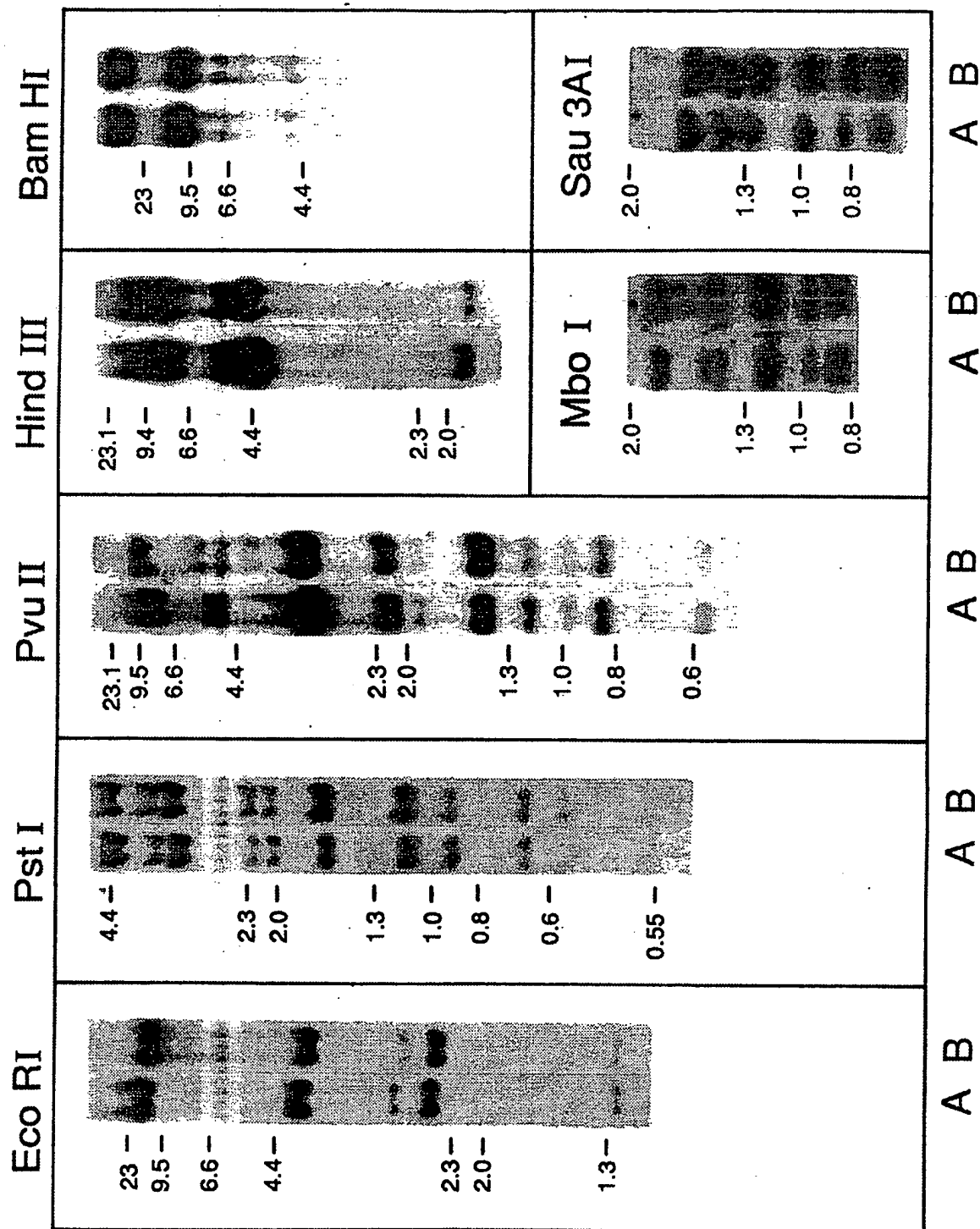


FIG. 2

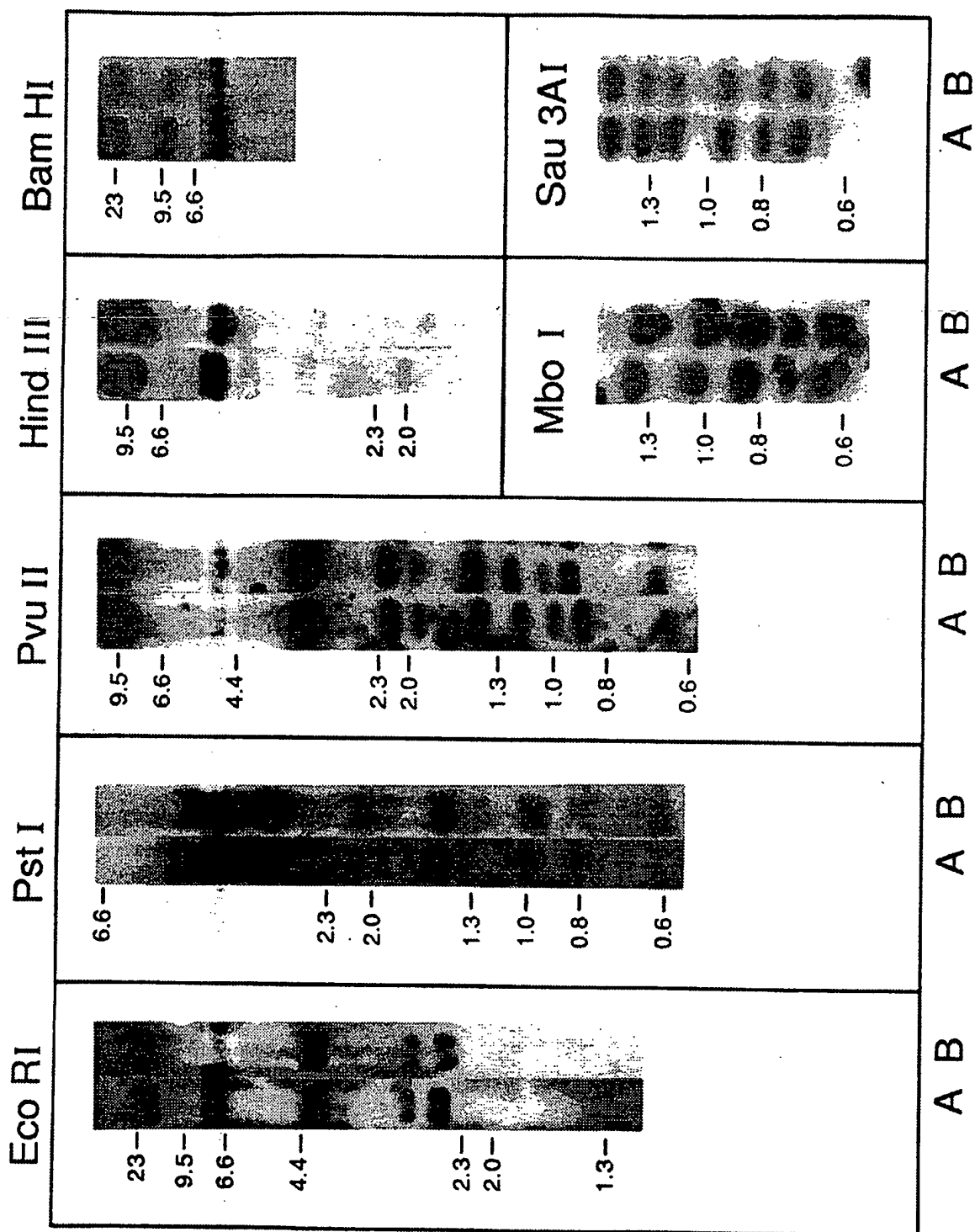


FIG. 3

FIG. 4

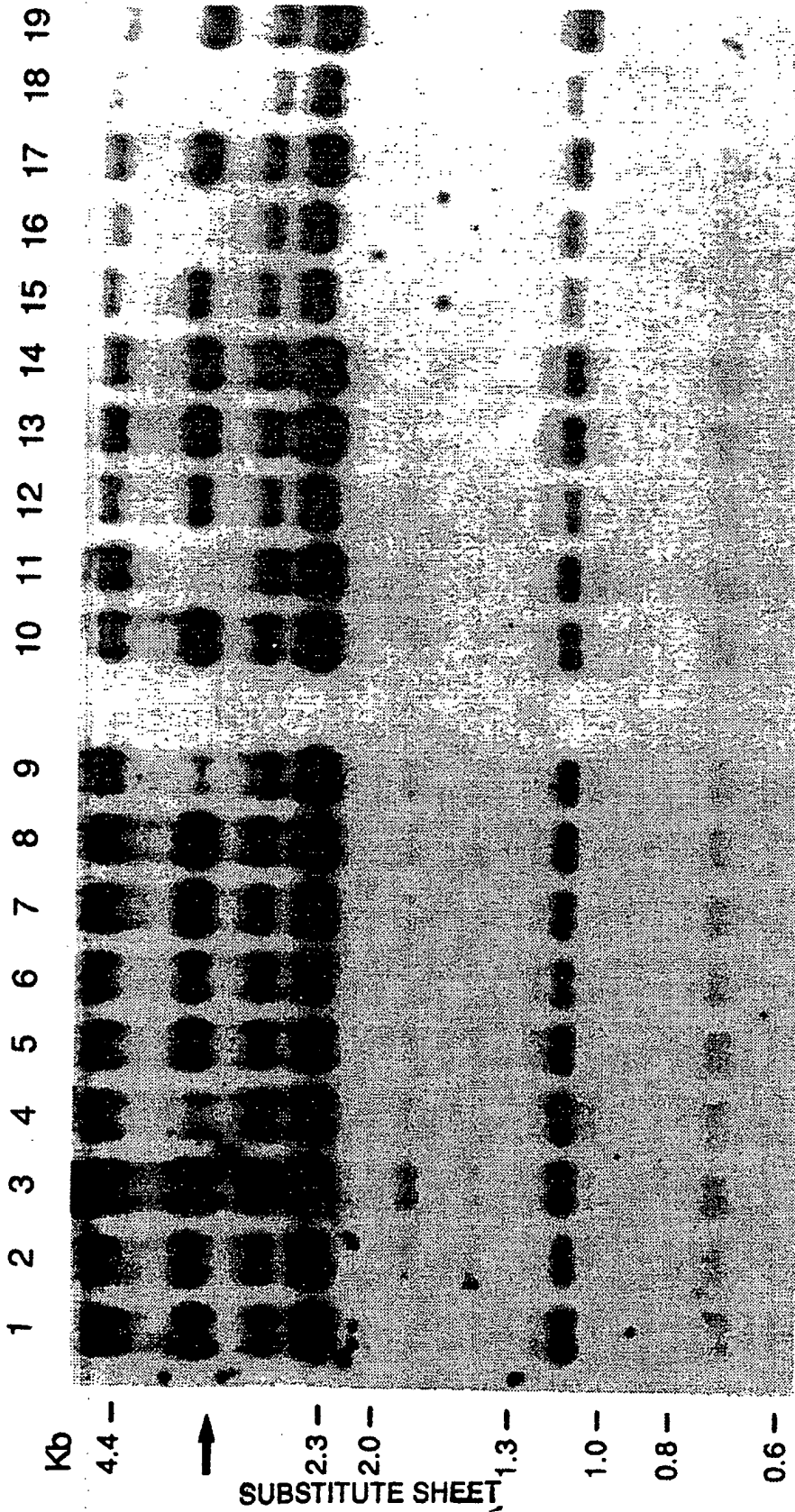
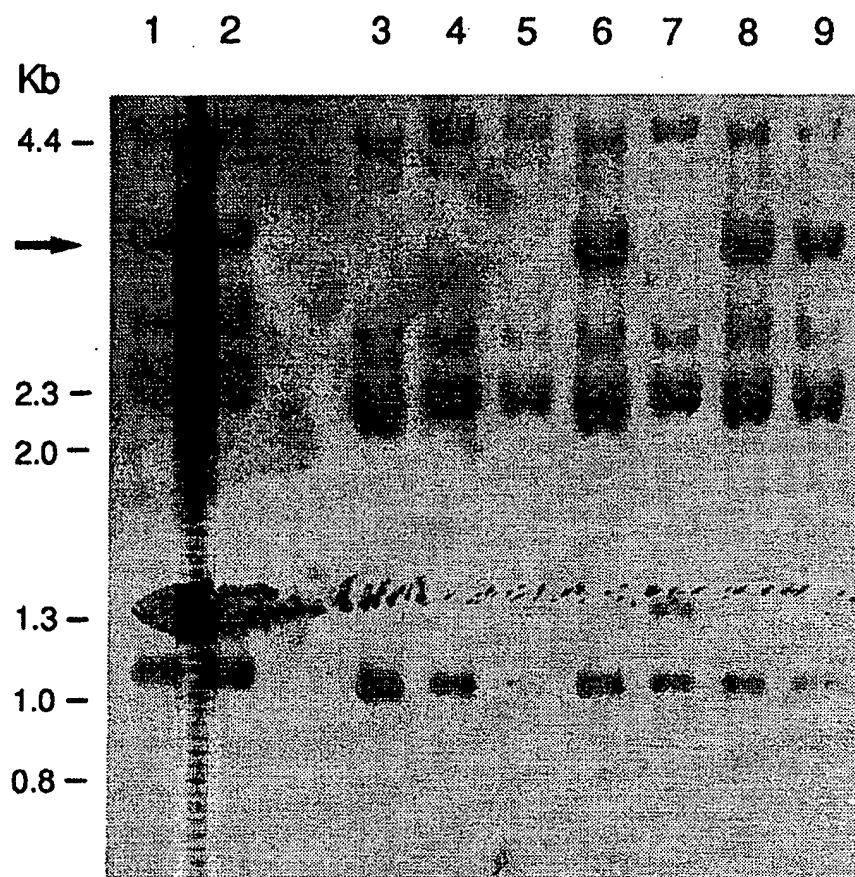


FIG. 5



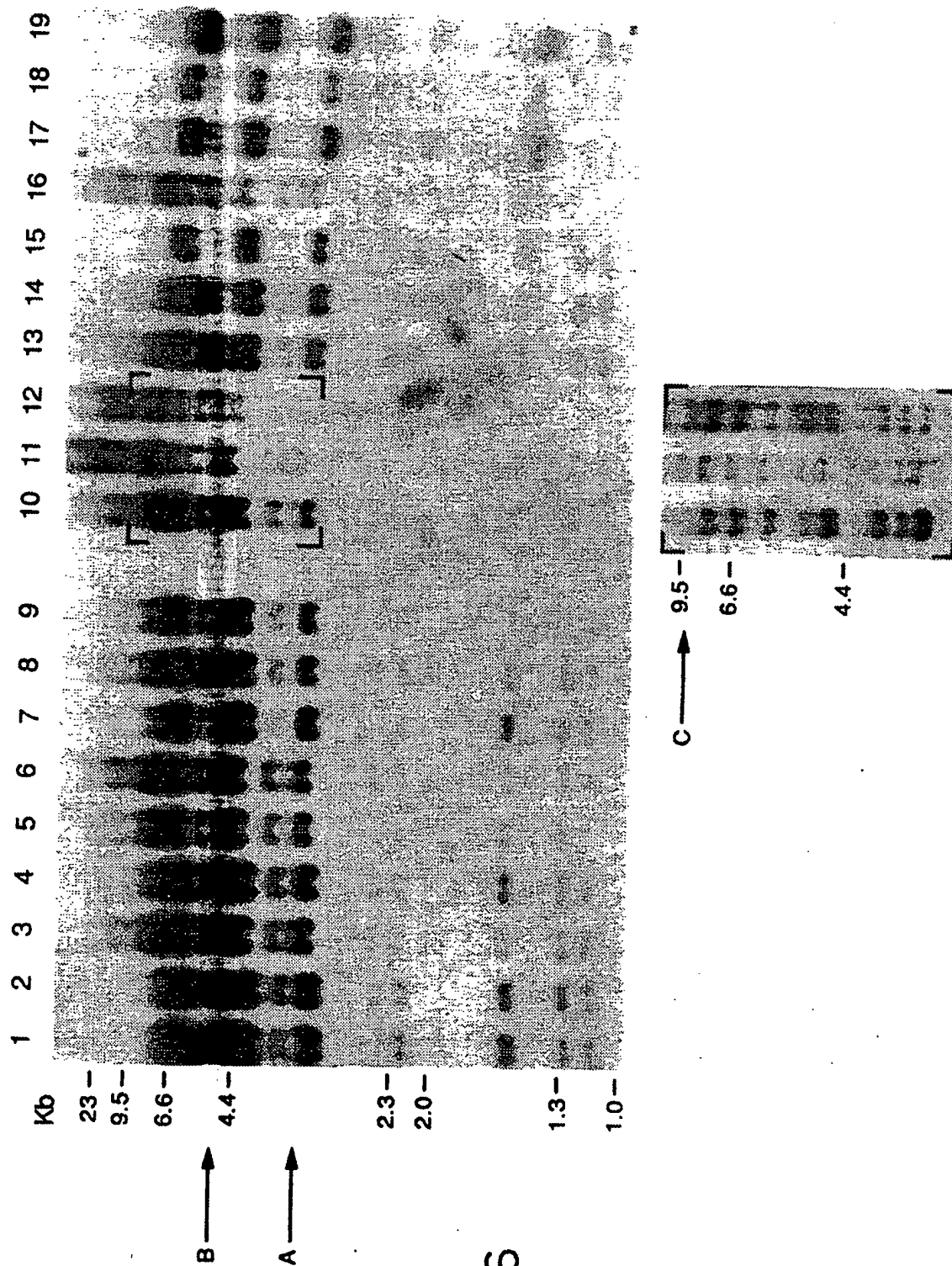


FIG. 6

FIG. 7

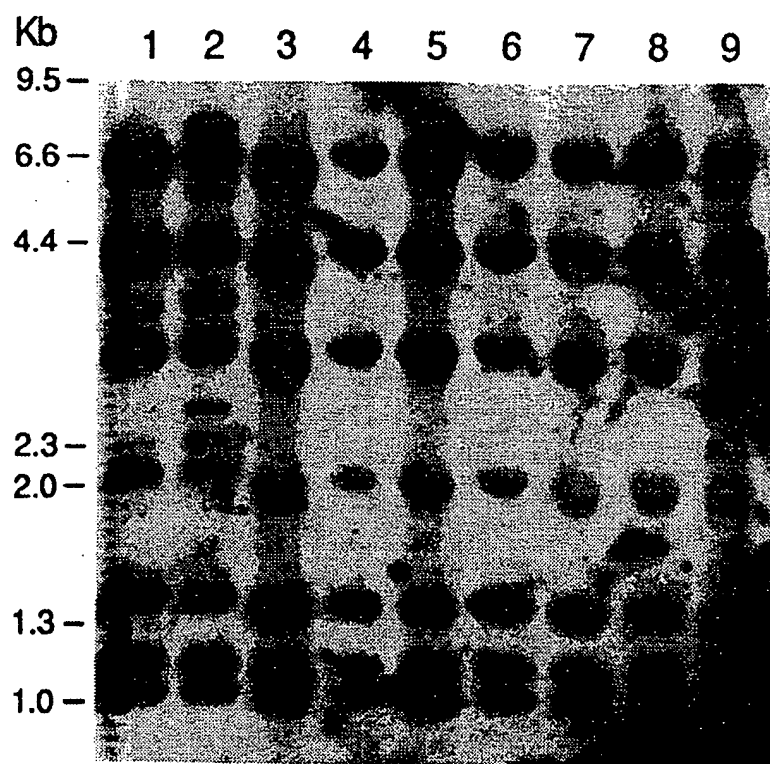
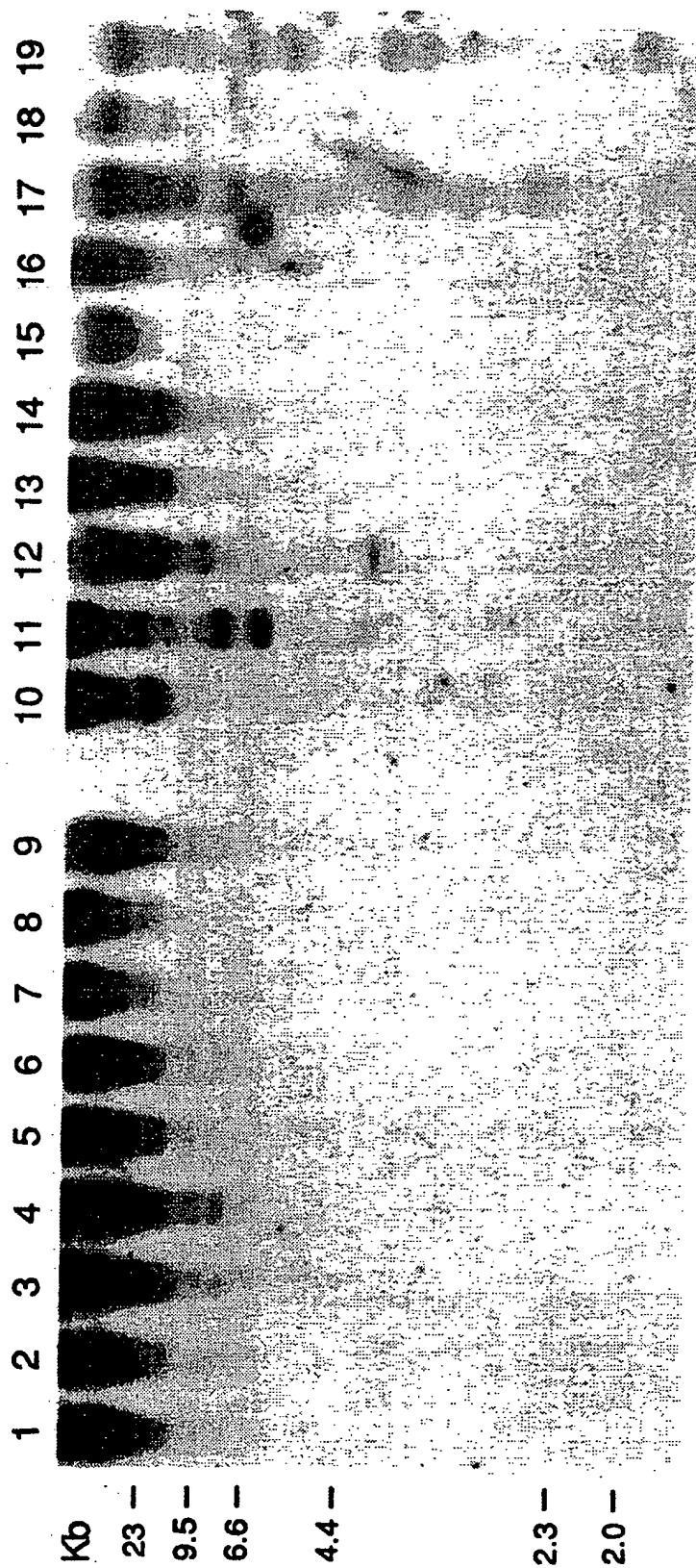


FIG. 8



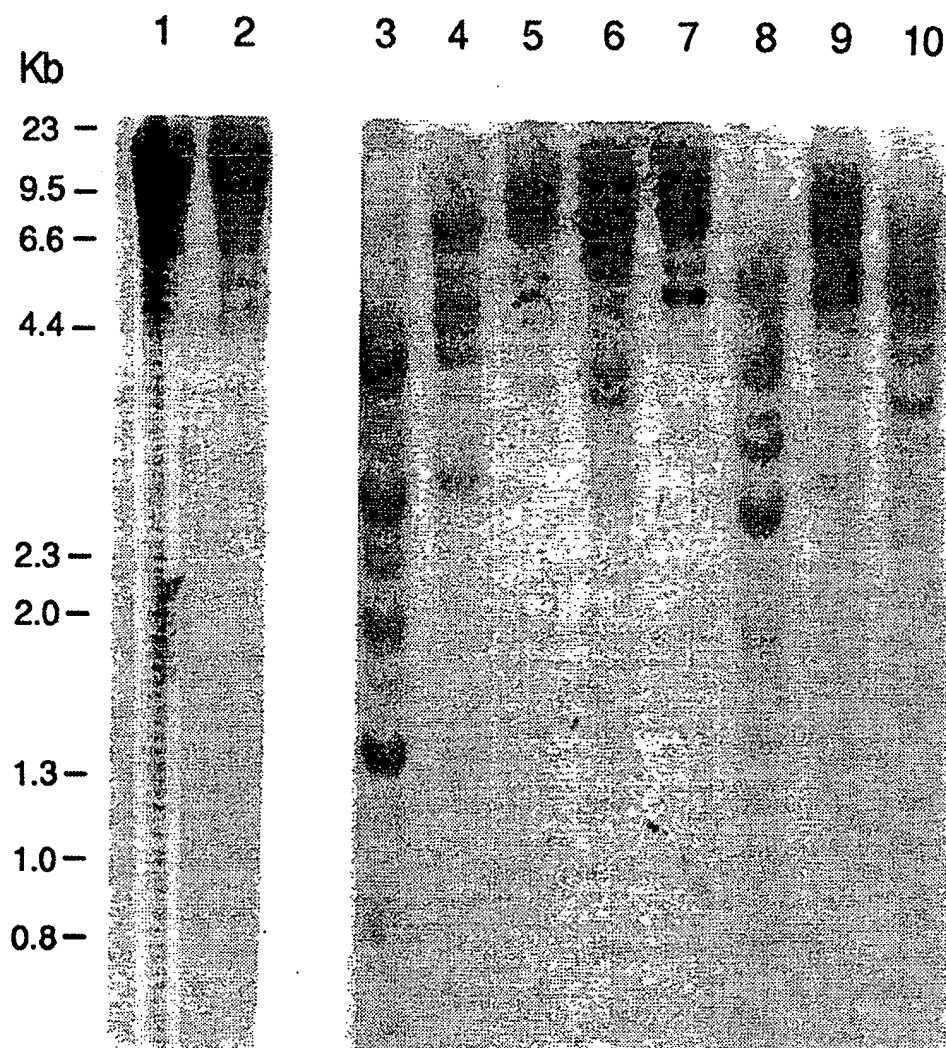


FIG. 9

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FIG. 10A

1 CTT GTC TTC CTC CTC CTC CTC GGG GCC CTC GGA CTG TGT CTG
 Leu Val Phe Leu Leu Phe Leu Gly Ala Leu Gly Leu Cys Leu
 GCT GGC CGT AGG
 Ala Gly Arg Arg

61 AGA AGG AGT GTT CAG TGG TGC GCC GTA TCC CAA CCC GAG GCC ACA AAA
 Arg Arg Ser Val Gln Trp Cys Ala Val Ser Gln Pro Glu Ala Thr Lys
 Asn
 TGC TTC CAA TGG
 Cys Phe Gln Trp

121 CAA AGG AAT ATG AGA AAA GTG CGT GGC CCT CCT GTC AGC TGC ATA AAG
 Gln Arg Asn Met Arg Lys Val Arg Gly Pro Pro Val Ser Cys Ile Lys
 Leu
 AGA GAC TCC CCC
 Arg Asp Ser Pro

181 ATC CAG TGT ATC CAG GCC ATT GCG GAA AAC AGG GCC GAT GCT GTG ACC
 Ile Gln Cys Ile Gln Ala Ile Ala Glu Asn Arg Ala Asp Ala Val Thr
 CTT GAT GGT GGT
 Leu Asp Gly Gly

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FIG. 10B

241 TTC ATA TAC GAG GCA GGC CTG GCC CCC TAC AAA CTG CGA CCT GTA GCG
 Phe Ile Tyr Glu Ala Gly Leu Ala Pro Tyr Lys Leu Arg Pro Val Ala
 GCG GAA GTC TAC
 Ala Glu Val Tyr

301 GGG ACC GAA AGA CAG CCA CGA ACT CAC TAT TAT GCC GTG GCT GTG GTG
 Gly Thr Glu Arg Gln Pro Arg Thr His Tyr Tyr Ala Val Ala Val Val
 AAG AAG GGC GGC
 Lys Lys Gly Gly

361 AGC TTT CAG CTG AAC GAA CTG CAA GGT CTG AAG TCC TGC CAC ACA GGC
 Ser Phe Gln Leu Asn Glu Leu Gln Gly Leu Lys Ser Cys His Thr Gly
 CTT CGC AGG ACC
 Leu Arg Arg Thr

421 GCT GGA TGG AAT GTC CCT ATA GGG ACA CTT CGT CCA TTC TTG AAT TGG
 Ala Gly Trp Asn Val Pro Ile Gly Thr Leu Arg Pro Phe Leu Asn Trp
 C
 Thr
 ACG GGT CCA CCT
 Thr Gly Pro Pro

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FIG. 10C

481 GAG CCC ATT GAG GCA GCT GTG GCC AGG TTC TTC TCA GCC AGC TGT GTT
 Glu Pro Ile Glu Ala Val Ala Arg Phe Phe Ser Ala Ser Cys Val
 CCC GGT GCA GAT
 Pro Gly Ala Asp

541 AAA GGA CAG TTC CCC AAC CTG TGT CGC CTG TGT GCG GGG ACA GGG GAA
 Lys Gly Gln Phe Pro Asn Leu Cys Arg Leu Cys Ala Gly Thr Gly Glu
 AAC AAA TGT GCC
 Asn Lys Cys Ala

601 TTC TCC TCC CAG GAA CCG TAC TTC AGC TAC TCT GGT GCC TTC AAG TGT
 Phe Ser Ser Gln Glu Pro Tyr Phe Ser Tyr Ser Gly Ala Phe Lys Cys
 CTG AGA GAC GGG
 Leu Arg Asp Gly
 Lys

661 GCT GGA GAC GTG GCT TTT ATC AGA GAG AGC ACA GTG TTT GAG GAC CTG
 Ala Gly Asp Val Ala Phe Ile Arg Glu Ser Thr Val Phe Glu Asp Leu
 TCA GAC GAG GCT
 Ser Asp Glu Ala

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FIG. 10D

721 GAA AGG GAC GAG TAT GAG TTA CTC TGC CCA GAC AAC ACT CGG AAG CCA
 Glu Arg Asp Glu Tyr Glu Leu Leu Cys Pro Asp Asn Thr Arg Lys Pro
 GTG GAC AAG TTC
 Val Asp Lys Phe

781 AAA GAC TGC CAT CTG GCC CGG GTC CCT TCT CAT GCC GTT GTG GCA CGA
 Lys Asp Cys His Leu Ala Arg Val Pro Ser His Ala Val Val Ala Arg
 AGT GTG AAT GGC
 Ser Val Asn Gly

841 AAG GAG GAT GCC ATC TGG AAT CTT CTC CGC CAG GCA GAA AAG TTT
 Lys Glu Asp Ala Ile Trp Asn Leu Leu Arg Gln Ala Gln Glu Lys Phe
 GGA AAG GAC AAG
 Gly Lys Asp Lys

901 TCA CCG AAA TTC CAG CTC TTT GGC TCC CCT AGT GGG CAG AAA GAT CTG
 Ser Pro Lys Phe Gln Leu Phe Gly Ser Pro Ser Gly Gln Lys Asp Leu
 CTG TTC AAG GAC
 Leu Phe Lys Asp

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FIG. 10E

961 TCT GCC ATT GGG TTT TCG AGG GTG CCC CCG AGG ATA GAT TCT GGG CTG
 Ser Ala Ile Gly Phe Ser Arg Val Pro Pro Arg Ile Asp Ser Gly Leu
 TAC CTT GGC TCC
 Tyr Leu Gly Ser

1021 GGC TAC TTC ACT GCC ATC CAG AAC TTG AGG AAA AGT GAG GAG GAA GTG
 Gly Tyr Phe Thr Ala Ile Gln Asn Leu Arg Lys Ser Glu Glu Val
 GCT GCC CGG CGT
 Ala Ala Arg Arg

1081 GCG CGG GTC GTG TGG TGT GCG GTG GGC GAG CAG GAG CTG CGC AAG TGT
 Ala Arg Val Val Trp Cys Ala Val Gly Glu Gln Glu Leu Arg Lys Cys
 AAC CAG TGG AGT
 Asn Gln Trp Ser

1141 GGC TTG AGC GAA GGC AGC GTG ACC TGC TCC TCG GCC TCC ACC ACA GAG
 Gly Leu Ser Glu Gly Ser Val Thr Cys Ser Ser Ala Ser Thr Thr Glu
 TAC CTT GGC TCC
 Tyr Leu Gly Ser

SUBSTITUTE SHEET

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FIG. 10F

1201 CTG GTG CTG AAA GGA GAA GCT GAT GCC ATG AGT TTT GAT GGA GGA TAT
 Leu Val Leu Lys Gly Glu Ala Asp Ala Met Ser Leu Asp Gly Gly Tyr

GTG TAC ACT GCA
 Val Tyr Thr Ala

1261 T GGC AAA TGT GGT TTG GTG CCT GTC CTG GCA GAG AAC TAC AAA TCC CAA
 Gly Lys Cys Gly Leu Val Pro Val Leu Ala Glu Asn Tyr Lys Ser Gln
 Cys

CAA AGC AGT GAC
 Gln Ser Ser Asp

1321 CCT GAT CCT AAC TGT GTG GAT AGA CCT GTG GAA GGA TAT CTT GCT GTG
 Pro Asp Pro Asn Cys Val Asp Arg Pro Val Glu Gly Tyr Leu Ala Val

GCG GTG GTT AGG
 Ala Val Val Arg

1381 AGA TCA GAC ACT AGC CTT ACC TGG AAC TCT GTG AAA GGC AAG AAG TCC
 Arg Ser Asp Thr Ser Leu Thr Trp Asn Ser Val Lys Gly Lys Lys Ser

TGC CAC ACC GCC
 Cys His Thr Ala

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FIG. 10G

1441	GTG GAC AGG ACT GCA GGC TGG AAT ATC CCC ATG GGC CTG CTC TTC AAC Val Asp Arg Thr Ala Gly Trp Asn Ile Pro Met Gly Leu Phe Asn
	GC Ala
	CAG ACG GGC TCC Gln Thr Gly Ser
1501	TGC AAA TTT GAT GAA TAT TTC AGT CAA AGC TGT GCC CCT GGG TCT GAC Cys Lys Phe Asp Glu Tyr Phe Ser Gln Ser Cys Ala Pro Gly Ser Asp
	CCG AGA TCT AAT Pro Arg Ser Asn
1561	CTC TGT GCT CTG TGT ATT GGC GAC GAG CAG GGT GAG AAT AAG TGC GTG Leu Cys Ala Leu Cys Ile Gly Asp Glu Gln Gly Glu Asn Lys Cys Val
	T CCC AAC AGC AAC Pro Asn Ser Asn
1621	GAG AGA TAC TAC GGC TAC ACT GGG GCT TTC CGG TGC CTG GCT GAG AAT Glu Arg Tyr Tyr Gly Tyr Thr Gly Ala Phe Arg Cys Leu Ala Glu Asn
	GCT GGA GAC GTT Ala Gly Asp Val

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FIG. 10H

1681 GCA TTT GTG AAA GAT GTC ACT GTC TTG CAG AAC ACT GAT GGA AAT AAC
 Ala Phe Val Lys Asp Val Thr Val Leu Gln Asn Thr Asp Gly Asn Asn
 AAT GAG GCA TGG
 Asn Glu Ala Trp

1741 GCT AAG GAT TTG AAG CTG GCA GAC TTT GCG CTG CTG TGC CTC GAT GGC
 Ala Lys Asp Leu Lys Leu Ala Asp Phe Ala Leu Cys Leu Asp Gly
 AAA CGG AAG CCT
 Lys Arg Lys Pro

1801 GTG ACT GAG GCT AGA AGC TGC CAT CTT GCC ATG GCC CCG AAT CAT GCC
 Val Thr Glu Ala Arg Ser Cys His Leu Ala Met Ala Pro Asn His Ala
 GTG GTG TCT CGG
 Val Val Ser Arg

1861 ATG GAT AAG GTG GAA CGC CTG AAA CAG GTG TTG CTC CAC CAA CAG GCT
 Met Asp Lys Val Glu Arg Leu Lys Gln Val Leu Leu His Gln Gln Ala
 AAA TTT GGG AGA
 Lys Phe Gly Arg

C

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FIG. 101

1921	AAT GGA TCT GAC TGC CCG GAC AAG TTT TGC TTA TTC CAG TCT GAA ACC Asn Gly Ser Asp Cys Pro Asp Lys Phe Cys Leu Phe Gln Ser Glu Thr
	AAA AAC CTT CTG Lys Asn Leu Leu
1981	TTC AAT GAC AAC ACT GAG TGT CTG GCC AGA CTC CAT GGC AAA ACA ACA Phe Asn Asp Asn Thr Glu Cys Leu Ala Arg Leu His Gly Lys Thr Thr
	TAT GAA AAA TAT Tyr Glu Lys Tyr
2041	TTG GGA CCA CAG TAT GTC GCA GGC ATT ACT AAT CTG AAA AAG TGC TCA Leu Gly Pro Gln Tyr Val Ala Gly Ile Thr Asn Leu Lys Lys Cys Ser
	C ACC TCC CCC TCC Thr Ser Pro Ser Leu
2101	TGG AAG CCT GTG AAT TC 2117 Trp Lys Pro Val Asn Leu Glu Ala Cys Glu Phe

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US92/04012

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :C12N 15/00, 15/10, 15/12; A61K 35/20

US CL :435/6, 69.1, 320.1; 514/6; 530/395, 400; 536/27

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/6, 69.1, 69.6, 320.1; 514/6; 530/350, 395, 400; 536/27

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS; MEDLINE, BIOSIS, World Patents Index

search terms: lactoferrin, gene, DNA, cDNA, breast cancer, cancer

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
<u>X</u> Y	Clinica Chimica Acta, Vol. 151, issued 1985, W.R. Bezwoda et al, "Enzyme linked immunosorbent assay for lactoferrin. Plasma and tissue measurements", pages 61-69, entire document.	<u>2,9</u> 1,3-8,10-11
<u>X</u> Y	Clinica Chimica Acta, Vol. 157, issued 1986, W.R. Bezwoda et al, "Isolation and characterisation of lactoferrin separate from human whey by adsorption chromatography using Cibacron Blue F3G-A linked affinity adsorbent", pages 89-94, entire document.	<u>2,9</u> 1,3-8
<u>X</u> Y	FEBS Letters, Vol. 109, no. 2, issued January 1980, L. Blackberg et al, "Isolation of lactoferrin from human whey by a single chromatographic step", pages 180-184, entire document.	<u>2,9</u> 1,3-8
Y	J. Sambrook et al., "Molecular cloning techniques, a laboratory manual", published 1989 by Cold Spring Harbor Laboratory Press, pages 12.2-12.15, entire document.	1,3-8

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be part of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

23 July 1992

Date of mailing of the international search report

31 JUL 1992

Name and mailing address of the ISA/
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

Authorized officer

DIAN COOK

Facsimile No. NOT APPLICABLE

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US92/04012

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Cancer Research, Vol. 46, no. 3, issued March 1986, K. Shirasuna et al, "Isolation and characterization of different clones including myoepithelial-like variants from a clonal neoplastic epithelial duct cell line of human salivary gland origin", pages 1418-1426, especially abstract.	12-14